Oral Administration of *Lactococcus lactis* Expressing Synthetic Genes of Myelin Antigens in Decreasing Experimental Autoimmune Encephalomyelitis in Rats

Kaja Kasarello, Barbara Kwiatkowska-Patzer, Andrzej W. Lipkowski, Jacek K. Bardowski, Agnieszka K. Szczepankowska

**Background:** Multiple sclerosis is a human autoimmunological disease that causes neurodegeneration. One of the potential ways to stop its development is induction of oral tolerance, whose effect lies in decreasing immune response to the fed antigen. It was shown in animal models that administration of specific epitopes of the three main myelin proteins – myelin oligodendrocyte glycoprotein (MOG), myelin basic protein (MBP), and proteolipid protein (PLP) – results in induction of oral tolerance and suppression of disease symptoms. Use of bacterial cells to produce and deliver antigens to gut mucosa seems to be an attractive method for oral tolerance induction in treatment of diseases with autoimmune background.

**Material/Methods:** Synthetic genes of MOG35-55, MBP85-97, and PLP139-151 myelin epitopes were generated and cloned in *Lactococcus lactis* under a CcpA-regulated promoter. The tolerogenic effect of bacterial preparations was tested on experimental autoimmune encephalomyelitis, which is the animal model of MS. EAE was induced in rats by intradermal injection of guinea pig spinal cord homogenate into hind paws.

**Results:** Rats were administered preparations containing whole-cell lysates of *L. lactis* producing myelin antigens using different feeding schemes. Our study demonstrates that 20-fold, but not 4-fold, intragastric administration of autoantigen-expressing *L. lactis* cells under specific conditions reduces the clinical symptoms of EAE in rats.

**Conclusions:** The present study evaluated the use of myelin antigens produced in *L. lactis* in inhibiting the onset of experimental autoimmune encephalomyelitis in rats. Obtained results indicate that application of such recombinant cells can be an attractive method of oral tolerance induction.

**MeSH Keywords:** Encephalomyelitis, Autoimmune • *Lactococcus lactis* • Myelin Proteins

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Background

Multiple sclerosis (MS) is an autoimmune disease that has a serious impact on physical abilities of the patient. It is postulated to involve cell-mediated and humoral responses directed against myelin proteins, including myelin basic protein (MBP), proteolipid protein (PLP), myelin-associated glycoprotein (MAG), and myelin oligodendrocyte glycoprotein (MOG) [1–6]. This immunoreactivity causes inflammation that leads to demyelination and axonal loss in the central nervous system (CNS) and results in slowing or even breaking neurotransmission [7]. The etiology of MS remains unknown and the commercially available medicines do not prevent disease development [8,9]. Currently, the most effective MS treatments are based on general immunosuppression or immunomodulation, which may lead to a number of adverse effects (e.g., common bacterial and viral infections, mood swings, high blood pressure, weight gain), including severe adverse effects such as progressive multifocal leukoencephalopathy. Data show that after 6 years of treatment, the disability of people with multiple sclerosis is the same as without drug therapy (Kwiatkowska-Patzer, personal communication).

An alternative treatment relies on induction of a state termed oral tolerance. Most simply, oral tolerance can be defined as the decrease of response to a previously fed antigen. It is a common physiological state preventing the formation of systemic immune responses to proteins ingested daily [10]. Preclinical trials show that on the same basis, oral administration of autoantigens induces a response suppressing experimental autoimmune diseases [11]. Studies using the experimental allergic encephalomyelitis (EAE), an animal model of MS, have shown that intradermal injection of myelin antigens such as MBP, MOG or PLP proteins with Freund’s adjuvant leads to development of acute paralysis [12–14]. However, oral administration of a myelin antigen or antigens before injection inhibits paralysis signs and immune cells infiltration as shown in histopathological studies [15,16]. Moreover, feeding with myelin antigens after the onset of EAE was determined to reduce disease development and facilitate remission [17]. Also, our own previous studies have demonstrated that intragastric administration of pig spinal cord hydrolysate, containing short peptide fragments of myelin proteins, induces oral tolerance in Wistar and Lewis rats with EAE [18–20]. Results indicated that the mixture of neuropeptides in the spinal cord hydrolysate given orally diminished immunological response to myelin antigens [21]. This initiated further studies on developing novel means of delivering myelin peptides to gut mucosa in combination with proficient production of selected autoantigens in bacterial cells.

Various studies show that peptides MOG35-55, MBP85-97, and PLP139-151 are epitopes that play a role in inhibiting autoantigen responses, both in MS patients and animals with EAE [17,22,23]. Therefore, they are generally considered as good antigens for oral tolerance induction. All of the above observations encourage continuing studies on the development of autoimmune disease treatment.

One of the strategies of delivering antigens to mucosal surfaces is the use of bacterial cells as carriers. Commonly, antigen delivery systems were based on attenuated pathogenic microorganisms or viruses such as *Salmonella typhi* or poliovirus [24,25]. An interesting alternative that has been gaining much interest in the last decades is the potential use of non-pathogenic bacteria as live oral vaccines. In this work we developed a system for production of selected epitopes from myelin proteins in a lactic acid bacterium (LAB). LAB are gram-positive, food-grade bacteria with various applications in food production and dairy fermentation [26,27]. Due to their recognized, non-pathogenic nature, LAB are of substantial interest for implementation in various research domains, including vaccine development, where they are considered as attractive antigen delivery vectors, and a good alternative to commonly used attenuated pathogens [28,29]. *Lactococcus lactis* is a model LAB that has been shown to efficiently produce heterologous proteins of different origin (e.g., viral, bacterial, mammalian) [30–33]. Numerous studies demonstrated that oral delivery of allergen- or autoantigen-expressing *L. lactis* cells can be a cost-effective and innovative means of inducing tolerance [34–36]. Among other evidence, it was shown that delivery of the ovalbumin-expressing *L. lactis* strain to the gut mucosal surface suppresses both systemic and local OVA-specific T-cell response [36].

These reports induced us to employ *L. lactis* as a host organism to produce myelin epitopes MOG35-55, MBP85-97, and PLP139-151 and deliver them to gut mucosa. For this, we generated synthetic genes of these peptide fragments and expressed them using the promoter region of the *L. lactis ptcB* gene (PptcB) inserted into the pIL253 vector. Previous studies have shown that the ptcB gene in *L. lactis* is engaged in sugar catabolism, and its promoter (PptcB) is regulated by CcpA, the catabolite control protein A [37]. The strength of PptcB was shown to vary depending on the sugar source in the medium. The highest promoter activation was determined to occur in the presence of cellobiose [37, Aleksandrzak-Piekarczyk – unpublished data]. Animal trials showed that oral administration of recombinant *L. lactis* cells encoding myelin epitopes can at certain doses inhibit progression of experimental autoimmune encephalomyelitis induced in rats. A scheme of antigen administration was also examined.
Table 1. Strains and plasmids.

<table>
<thead>
<tr>
<th>Strain</th>
<th>Genotype</th>
<th>Source</th>
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<tbody>
<tr>
<td>Lactococcus lactis IBB360</td>
<td>Natural isolate</td>
<td>IBB PAS collection</td>
</tr>
</tbody>
</table>

Plasmid

<table>
<thead>
<tr>
<th>Plasmid</th>
<th>Source</th>
</tr>
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<tbody>
<tr>
<td>pIL253:PptcB</td>
<td>Ery±, pIL253-derivative with ptcB gene promoter region</td>
</tr>
</tbody>
</table>

Material and Methods

Bacterial strains and plasmids

Strains and plasmids used in this work are listed in Table 1. Plasmid vector pIL253:PptcB, carrying the promoter region of L. lactis ptcB gene, was used in cloning procedures. L. lactis cells were grown at 30°C in M17 liquid medium supplemented with 0.5% cellobiose or on M17 solid medium containing 0.5% glucose [38]. Solid plate media contained 1.5% agar. When necessary the growth medium was supplemented with erythromycin at 5 µg/ml (for plasmid-carrying L. lactis derivatives).

DNA manipulations, transformation and sequencing

Standard DNA manipulations and cloning procedures were performed as described previously [39]. Transformation of L. lactis cells via electroporation was done as described elsewhere [40,41]. Digestions with restriction enzymes (Fermentas) were done according to the manufacturer’s instructions. DNA sequencing was done using a Big Dye sequencing kit. Sequences were analyzed with the BLAST program [42].

Cloning of MOG35-55, MBP85-97 and PLP139-151 synthetic gene fragments in L. lactis

Oligonucleotides used in this study are listed in Table 2. Two ‘LONG’ complementary oligonucleotides served as a template for amplification of neuropeptide-encoding sequences. Oligonucleotides were designed in accordance with the codon usage of L. lactis species to give the nucleotide sequence corresponding to the amino acid sequence of a given peptide fragment that is the most optimal for these bacteria. PCR reaction was performed with the Thermocycler apparatus using ExTaq DNA polymerase (TaKaRa) and, specific for each peptide, ‘SHORT’ primer pairs complementary to the 5’ ends of ‘LONG’ primers (Table 2). The resulting PCR products were then digested with PstI and SalI restriction enzymes (Fermentas) and ligated into the pIL253:PptcB vector cut with the same pair of enzymes. Ligated molecules were electroporated into L. lactis cells. Transformants were selected on M17 medium (Oxoid) supplemented with 1.5% agar, 0.5% glucose, and 5 µg/ml erythromycin to isolate cells carrying either the empty [pIL253:PptcB] or recombinant vectors. Colonies carrying single neuropeptide-containing plasmids were analyzed by ‘colony PCR’ technique using specific primers: ptcBfor/pILrev (Table 2). From confirmed proper recombinant cells, plasmid DNA was isolated using the Plasmid Mini Kit (A&A Biotechnology) and subjected to restriction analysis with HindIII enzyme (Fermentas). Obtained digestion pattern was compared to the pattern generated using the CloneManager 9 (Sci-Ed Software) program. Finally, nucleotide sequences of the cloned DNA fragments were examined for conformity with the nucleotide sequences of the synthetic myelin genes by DNA sequencing technique.

Transcriptional analysis of MOG35-55, MBP85-97 and PLP139-151 synthetic gene expression in L. lactis

Expression of the cloned neuropeptide synthetic gene sequences was analyzed by RT-PCR using SuperScriptIII Invitrogen). Total RNA was isolated from recombinant L. lactis cells carrying individual neuropeptide synthetic genes using High Pure RNA isolation kit (Roche). To eliminate residual DNA, total RNA was additionally digested with DNase I (Sigma) and subjected to RT-PCR reaction using a reverse ‘SHORT’ primer specific for each neuropeptide-encoding gene (Table 2).

Preparation of recombinant L. lactis whole-cell extracts

Recombinant lactococci were prepared for intragastric administration as follows. After o/n incubation at 30°C, the bacterial culture was harvested (8 000 g, 10 min., 4°C) and washed once with 0.9% NaCl. Then, cells were suspended in 0.9% NaCl and disrupted 3 times for 1 min using the MiniBeadbeater (BioSpec Products) and 106-µm glass beads (Sigma). To determine the amount of myelin peptides that should be delivered to the gut mucosa to evoke tolerance, adequate dilutions of cell lysates, corresponding to doses 10–106 CFU (Colony Forming Units) per 0.5 ml were made. Single doses were frozen in Eppendorf tubes in liquid nitrogen and stored at –20°C until needed.

Tolerance induction in rats with EAE using whole-cell extracts of L. lactis producing MOG35-55, MBP85-97 and PLP139-151

Female Lewis rats, 180–200 g, were fed intragastrically with a gauged pointed needle, four times during one week (every...
second day) or once a day for 20 consecutive days (from day −10 to +9). Preparations given to rats contained whole-cell L. lactis lysates diluted in 0.5 ml PBS accordingly to give $10^{-10}$–$10^{-8}$ CFU per feeding. EAE was evoked a week after the last feeding in respect to the 4-day scheme or on day 10 of the 20-day schedule. The day of immunization is marked as 0 DPI – day post immunization. To induce EAE, rats received intradermal injection of 50% guinea pig spinal cord homogenate in PBS, with Freund’s adjuvant (1:1) and Mycobacterium tuberculosis (4 mg/ml), into hind paws (100 µl/paw). Rats were weighed every day and clinical symptoms were evaluated from the day they first occurred, using a 5-grade scale: 1 – limp tail, 2 – hind leg weakness, 3 – hind leg paralysis and incontinence, 4 – front and hind leg weakness or quadriplegia, 5 – death. On day 14 after immunization (DPI) animals were sacrificed.

Results

Designing of synthetic MOG35-55, MBP85-97 and PLP139-151 gene sequences

Synthetic genes encoding human-derived MOG35-55 (from MOG protein; gi: 56388814) MBP85-97 (from MBP protein; gi: 56388814) and PLP139-151 (from PLP protein; gi: 56388814) were designed according to the published sequences.

### Table 2. Oligonucleotide sequences used in study.

<table>
<thead>
<tr>
<th>Neuropeptide</th>
<th>Oligonucleotide (LONG) used as DNA templates for amplification of synthetic neuropeptide genes</th>
<th>Oligonucleotide (SHORT) used as primers for amplification of synthetic neuropeptide genes</th>
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</thead>
<tbody>
<tr>
<td>MOG35-55</td>
<td>FLONGMOG35 5' GTATTTCTATGGAAGTTGGATGGTATCGTTCACC AATTTCACCGTTGTCTTTATGTAATGG 3'</td>
<td>RSHORTMOG35 5' AAAGTGGATATTTATTTTCCATTACGATA 3'</td>
</tr>
<tr>
<td></td>
<td>RLONGMOG35 5' CCATTAGCATATAAGTAAAGCAGTGAAATGCTTCAAGTAAAAAT 3'</td>
<td></td>
</tr>
<tr>
<td>MBP85-97</td>
<td>FLONGMBP85 5' GTATTTCTATGCCAGGATCACGTCCACATTTAATTCGTTTATTTTCACGT 3'</td>
<td>RSHORTMBP85 5' AAAGTGGATATTTATTTTCCATTACGATA 3'</td>
</tr>
<tr>
<td></td>
<td>RLONGMBP85 5' ACCTGGAAAAATACGTTACGTTTCCTAATGAATGCATAGAAATAC 3'</td>
<td></td>
</tr>
<tr>
<td>PLP139-151</td>
<td>FLONGPLP139 5' GTATTTCTATGCATTCATTAGGAAAATGGTTAGGACATCCAGATAAATTT 3'</td>
<td>RSHORTPLP139 5' AAAGTGGATATTTATTTTCCATTACGATA 3'</td>
</tr>
<tr>
<td></td>
<td>RLONGPLP139 5' AAAGTGGATATTTATTTTCCATTACGATA 3'</td>
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</table>

* RBS and spacer sequence marked in bold, sequences recognized by restriction enzymes are underlined.

Analysis of the therapeutic effect of L. lactis-produced peptide preparations on rats

The efficiency of bacterial preparations were assessed by calculating (i) mean score of clinical symptoms, and (ii) body mass, reflecting the overall condition of the animal. First clinical symptoms of EAE were observed at the 11 DPI. Peak of clinical symptoms was noted between 11–14 DPI. The mean clinical score was calculated for this time range. Body mass index was counted as the ratio of body mass at the end of experiment [14 DPI] to body mass at the day of immunization [0 DPI]. Body mass at 0 DPI was considered as 1. Statistical evaluations were made using the Mann-Whitney test. A value of p less than 0.05 was considered as statistically significant.
gi: 54038405AAH84713.1) and PLP139-151 (from PLP protein; gi: 13591880) epitopes were obtained by PCR method. For each peptide fragment, two complementary oligonucleotides (for/rev ‘LONG’) were used as template. To ensure efficient gene expression associated with faster translation rates, all ‘LONG’ primers used in this study were designed taking into account optimal codon usage in L. lactis, obtained from the Codon Usage Database (www.kazusa.or.jp). Additionally, ‘LONG’ primers contained at their 5’ ends sequences specific for the Lactococcus lactis RBS (ribosome-binding site; AAGGAG) consensus sequence recognized by the translation machinery and ‘spacer’ sequence (TATTTCT) localized between the RBS region and the translation START codon. The forward ‘LONG’ primer used to amplify the PLP139-151-encoding gene was modified by introducing a sequence corresponding to the translation START codon (ATG), just before the original peptide sequence. Genes encoding myelin peptide fragments were generated by two oligonucleotides (for/rev ‘SHORT’) homologous to the extremities of the ‘LONG’ primers (Table 2).

Cloning of MOG35-55-, MBP85-97- and PLP139-151-encoding genes in L. lactis

The generated synthetic genes were cloned separately into L. lactis-replicating pIL253:PptcB plasmid. Obtained recombinant vectors were introduced independently into electrocompetent cells by using an efficient transformation technique commonly used for L. lactis [40,41]. Clones containing synthetic genes were identified in the culture population by analyzing the obtained transformants using ‘colony PCR’ technique for the presence of inserts corresponding in length to the expected DNA fragments (Figure 1). PCR performed directly on grown cells using specific primers, allowed isolating colonies specifically carrying recombinant vectors. Digestion patterns of obtained from recombinant plasmid DNA isolated from these colonies were in conformity with the predicted restriction pattern generated using bioinformatics program (Figure 2). Such result, in parallel with DNA sequencing, provided proof that these plasmids do not undergo any rearrangements and are stably maintained in L. lactis, suggesting at the same time that the encoded myelin peptides are not toxic to the cells.

Figure 1. Colony PCR on selected transformant clones. (A) pIL253:PptcB:MOG35-55 (8 clones tested), (B) pIL253:PptcB:MBP85-97 (3 clones tested), (C) pIL253:PptcB:PLP139-151 (12 clones tested). Colony PCR carried out on transformant clones obtained after electroporation of L. lactis cells with recombinant constructs and selection of M17 solid medium supplemented with 0.5% glucose and erythromycin at 5 µg/ml. Specific primers used (ptcBfor/pILrev) were homologous to the vector sequence flanking the potentially cloned fragment. Expected product size: ~550 bp (marked with red arrows), 1 kb DNA Ladder (Fermentas) used as DNA size reference marker.

Figure 2. Control digestions of recombinant plasmid DNA using HindIII (Fermentas) enzyme. (A) pIL253:PptcB:MOG35-55, (B) pIL253:PptcB:MBP85-97, (C) pIL253:PptcB:PLP139-151. Expected DNA fragments after restriction analysis for correct constructs: 3897 bp, 845 bp (marked by red arrows), 1 kb DNA ladder (M) 1kb DNA Ladder (Fermentas) used as DNA size reference marker.
MOG35-55 fragments corresponding to doses 10^3 (n=6) were fed for 20 consecutive days with whole-cell lysates of recombinant (n=6) L. lactis. To determine the optimal amount of myelin peptides for effective induction of oral tolerance, animals with evoked EAE (n=6) were fed for 20 consecutive days with whole-cell lysates of recombinant L. lactis producing MBP85-97, PLP139-151 and MOG35-55 fragments corresponding to doses 10^3–10^6 CFU –10^3 CFU in comparison to non-fed EAE animals (Figure 5B). To examine whether the cloned synthetic neuropeptide genes are properly expressed in L. lactis, transcriptional studies using RT-PCR have been carried out. Total RNA was isolated from bacterial strains carrying the recombinant plasmids and, after specific treatment, was subjected to reverse transcription using a specific reverse ‘SHORT’ primer complementary to the 3’ end of the strand encoding the particular synthetic gene (Table 2). Obtained cDNAs were subsequently subjected to amplification by the classical PCR technique using two ‘SHORT’ primer pairs (for/rev) homologous to each neuropeptide sequence (Table 2). In result, DNA fragments were obtained which length corresponded to the length of individual synthetic genes confirming the presence of synthetic neuropeptide gene transcripts (Figure 3).

Transcriptional studies on the expression of synthetic MOG35-55, MBP85-97 and PLP139-151 genes in L. lactis

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Preliminary determination of optimal dose of L. lactis-produced myelin peptides for oral tolerance induction

In first place, we concentrated our efforts on choosing the proper dose of recombinant L. lactis bacteria, which would elicit the desired effect, which is a crucial step in obtaining tolerance. We applied two simple parameters: (i) mean score, which is widely used to reflect the physical condition of the animals and (ii) body mass of the animals, which allows assessing the general state of the animals (it is clear that the body mass of the animal with ongoing inflammatory process will drop).

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Results obtained from this preliminary trial show influence of certain doses on EAE progression in rats. Feeding with bacterial lysates corresponding to doses, 10^3, 10^4 and 10^6 CFU exhibited the tendency to decrease the mean score, although this was not statistically relevant (Figure 4A). Significant (p<0.001) body mass reduction was observed in EAE rats in comparison to intact animals (NT) (Figure 4B). Application of bacterial preparations showed a slight trend to increase the body mass of immunized animals, but this effect was statistically relevant (p<0.05) only in respect to dose 10^6 CFU. However, the same dose was ineffective in declining EAE progression. Thus, for further experiments two doses, 10^3 and 10^6 CFU, were selected, which led at the same time to both, the decrease of mean score and increase of body mass vs. non-fed EAE animals.

Induction of oral tolerance by L. lactis-produced myelin peptides

To confirm that the chosen bacterial doses (10^3 and 10^6 CFU) are efficient in inhibiting the on-set of EAE, a subsequent experiment was set-up on a larger animal group (n=12) using a 20-day feeding scheme. Results confirmed our previous observations. Both doses (10^3, 10^6) did not only significantly decrease (p<0.05) clinical symptoms (Figure 5A), but also reduced body mass drop in fed vs. non-fed EAE animals (Figure 5B). To investigate whether the observed effect is caused by myelin peptide fragments and not by bacteria themselves, whole-cell lysates of L. lactis containing the empty plasmid (pIL253:PptcB) administered at the same doses (C 10^3, C 10^6) were used as control. No relevant differences in the mean score compared to non-fed EAE animals were observed (Figure 5A). In contrast, there was a statistically significant increase in the body mass index observed for animals fed with the control preparation at dose 10^6 CFU in comparison to non-fed EAE animals (Figure 5B). Still, the observed effect was lower than for myelin-producing bacteria.

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symptoms in fed vs. non-fed EAE animals (Figure 6A). Body mass index was higher only for animals fed with preparations corresponding to $10^6$ CFU/dose (Figure 6B). Also for this group a slight decrease of clinical symptoms was observed (Figure 6A).

We also did not note any dissimilarities in the mean score between fed and non-fed animals when lysates of *L. lactis* containing the empty plasmid were applied in 4-day feeding regimen (Figure 6A). Similarly as in the 20-fold feeding scheme, a substantial variation of body mass index was noted for animals fed with the control preparation at dose $10^6$ CFU in comparison to non-fed EAE animals (Figure 6B).

**Figure 4.** Animals with EAE fed once per day for 20 consecutive days with preparations containing mixed whole-cell lysates of recombinant *L. lactis* strains at doses $10^1$–$10^8$ CFU (groups $10^1$–$10^8$), producing MBP85-97, PLP139-151 and MOG35-55 fragments. NT – non-treated animals, EAE – non-fed animals with EAE. (A) Mean score calculated at the peak of clinical symptoms [11–14 DPI]. n=6. (B) Body mass index at 14 DPI. Body mass at 0 DPI considered as 1. n=6. * p<0.05; *** p<0.001.

**Figure 5.** Animals with EAE fed once per day for 20 consecutive days with preparations containing mixed whole-cell lysates of recombinant *L. lactis* cells in doses $10^3$ or $10^6$ CFU, containing the empty plasmid pIL253:PptcB (groups C $10^3$ and C $10^6$) and producing MBP85-97, PLP139-151 and MOG35-55 fragments (groups $10^3$ and $10^6$). NT – non-treated animals, EAE – non-fed animals with EAE. (A) Mean score calculated at the peak of clinical symptoms [11–14 DPI]. n=12. * p<0.05. (B) Body mass index at 14 DPI. Body mass at 0 DPI considered as 1. n=12. * p<0.05; ** p<0.01; *** p<0.001.
ANIMAL STUDY

years have demonstrated that production of heterologous status by the American Food and Drug Administration. Recent tion has been assigned the GRAS (generally recognized as safe) due to its long and safe use in human and animal food produc Lactococcus lactis approach for induction of specific tolerance [36,47,48]. has been proven by multiple studies as a promising and effective es by using live microbial vectors as antigen carriers has also subunit) and other [43–46]. Treatment of autoimmune diseas antigens with mucosa-binding molecules (e.g., cholera toxin B tigen administration enclosed in microsphere beads, fusion of 5. These facts inclined us to investigate the potential ap myelin epitopes of the mammalian central nervous system and myelin proteins of the central nervous system [57]. In this study we have synthetically generated genes encoding myelin epitopes of the mammalian central nervous system and cloned them in L. lactis cells to assess their potential application in specific induction of oral tolerance. To express myelin peptide genes we used a pIL253-derived plasmid vector containing the L. lactis ptcB promoter. Our unpublished studies show that activity of the CcpA-controlled PptcB region, both plasmid- or chromosomally-located, is higher on cellulose (activator) than on glucose due to catabolite repression [37,58, Aleksandrzak-Piekarczyk – unpublished data]. Therefore, to ensure efficient proteins in L. lactis for vaccine development is efficient and economically substantial [33,49–51]. A great advantage of using Lactococcus in medical applications is that it is non-pathogenic and, as a Gram-positive bacteria, does not produce endotoxic lipopolysaccharides, and finally, does not induce inflammatory responses in healthy individuals. Oral administration of L. lactis expressing antigens or allergens was previously shown to be proficient in induction of mucosal immune responses [52–54]. Moreover, L. lactis cells were deter mined to exhibit an adjuvant effect when administered as live antigen delivery vectors or a peptidoglycan matrix component [55,56].

Discussion

One of the alternative means of treating MS is induction of oral tolerance against natural myelin proteins, such as MBP, MOG, PLP, which according to the current knowledge are engaged in MS pathophysiology and its animal model – EAE [12–14]. Studies show that administration of specific myelin peptides or whole mammalian spinal cord hydrolysates (e.g., from pig, rat, mouse) prevents or partially inhibits EAE progression [15,16,18–21]. However, free antigen delivery for therapeutic purposes has several drawbacks, including laborious procedures in achieving adequate protein quantity and purity. Implementation in clinical therapy of novel, efficient immunomodulation strategies based on autoantigen presentation, in first place, aims at: (i) production of sufficient amounts of autoantigens, and (ii) simple and stable delivery of antigens to mucosal surfaces. Taking this into account, various antigen-presenting systems have been developed, including antigen production in transgenic plants or antigen administration enclosed in microsphere beads, fusion of antigens with mucosa-binding molecules (e.g., cholera toxin B subunit) and other [43–46]. Treatment of autoimmune diseases by using live microbial vectors as antigen carriers has also been proven by multiple studies as a promising and effective approach for induction of specific tolerance [36,47,48].

Lactococcus lactis is a food-grade commensal bacterium, which due to its long and safe use in human and animal food production has been assigned the GRAS (generally recognized as safe) status by the American Food and Drug Administration. Recent years have demonstrated that production of heterologous

Figure 6. Animals with EAE fed once per day for 4 days within a week with preparations containing mixed whole-cell lysates of recombinant L. lactis strains in doses 10^3 or 10^6 CFU, containing the empty plasmid pIL253:PptcB (groups C 10^3 and C 10^6) and producing MBP85-97, PLP139-151 and MOG35-55 fragments (groups 10^3 and 10^6). NT – non-treated animals, EAE – non-fed animals with EAE.

(A) Mean score calculated at the peak of clinical symptoms [11-14 DPI]. n=9. (B) Body mass index at 14 DPI. Body mass as 0 DPI considered as 1. n=9.

* p<0.05; ** p<0.01; *** p<0.001.
peptide production, cells were grown in medium with cellobiose. Based on two parameters, mean score and body mass, our results provide evidence that preparations of recombinant *L. lactis* encoding myelin peptides grown on medium with cellobiose render a tolerogenic effect when administered orally to EAE rats.

In the study, EAE was evoked using guinea pig spinal cord homogenate, successfully used in previous studies [21]. By using the mixture of all myelin antigens, we imitate MS, where a broad spectrum of autoantigens is believed to be engaged in pathology. Application of an antigen mix containing three myelin epitopes was expected to more surely guarantee EAE suppression than a single antigen. Moreover, a mix of antigens resembles more closely the natural, physiological situation of tolerance induction in the gut. Our study shows that use of the microbially-expressed myelin epitopes, MOG35-55, PLP139-151, MBP85-97, induces oral tolerance in rats. Although, there is a limited number of works that examined the tolerogenic effect of peptide mixes, most recently Juryńczyk et al. [23] showed that transdermal application of a mix of the same peptides can be a promising approach in treatment of MS patients.

Among the crucial factors in obtaining the desired effect of oral tolerance are optimal antigen dose and feeding regimen. Our own previous experiments showed that administration of pig spinal cord hydrolysate based on a 4-day regimen during 1 week successfully induced oral tolerance [21]. The second (20-day) scheme was implemented in experiments on recombinant lactobacilli producing myelin proteins [47]. In this work we aimed at determining the most favorable feeding scheme for the designed lactococcal recombinant cells. Results from conducted trials showed that feeding EAE animals according to a 20-day scheme with a mixture of whole-cell lysates of *L. lactis* cells producing MBP85-97, PLP139-151 or MOG35-55, at 10^4 or 10^6 CFU/dose, is sufficient in reducing clinical symptoms and decreasing further body mass drop in fed vs. non-fed EAE animals. The fact that the same effect was obtained by applying these two concentrations could be explained by the dose-dependent mechanism of oral tolerance development, which occurs via clonal anergy or deletion (high doses), or active suppression (low doses) [11,59].

In contrast, the 4-fold feeding regimen did not reduce the clinical symptoms of EAE. This may be owed to the necessity to precisely determine the dose of *L. lactis* administered to animals that would render the expected effect. Performing a pilot test using a range of doses (10^3–10^6 CFU/dose), similarly to the one made for the 20-day feeding scheme, might be useful in determining whether other bacterial doses than 10^4 and 10^6 CFU are more efficient in the 4-time feeding procedure. However, it seems that longer exposure of autoantigens in the gut resembles more the natural feeding scheme, implied to be more advantageous than short-term feedings [60].

Another significant factor that should to be assessed is the time-scheme for antigen administration. Oral tolerance can be induced both, before or after disease induction, depending on the conditions of study (e.g., antigen dose and disease progression). Yet, numerous studies report that better suppression is observed when the antigen is delivered before immunization [61–64]. Feeding animals after immunization was reported to render limited suppression; yet, a prolonged feeding regimen was necessary [61,65]. Therefore, in our study animals were exposed to the recombinant antigen for 10 instead of 4 days before immunization and additionally for 10 days after immunization to assure better oral tolerance induction. Intragastric administration of *L. lactis* containing the empty plasmid was also determined to influence the overall condition of EAE-induced rats. Although body mass gain/loss is not a direct parameter associated with oral tolerance, such effect could indicate the regulatory property of *L. lactis* bacteria. Literature data suggests that *Lactococcus* could increase tolerogenic signals induced by delivered autoantigens by influencing: (i) processing or presentation of the antigen, and/or (ii) expression of co-stimulatory molecules on dendritic cells by affecting natural immune molecules [56]. In contrast, clinical symptoms in rats fed with control bacteria lysates were not as significantly inhibited in comparison to rats fed with lysates of recombinant bacteria encoding myelin epitopes, and only slightly lower than for non-fed EAE rats. This suggests that although *L. lactis* cells alone can show some tolerogenic effect in the mucosa, myelin antigens are necessary in decreasing the clinical symptoms of EAE. The perspective of using *L. lactis* as a carrier may (only) play a boosting effect in evoking oral tolerance.

**Conclusions**

In summary, we synthetically generated genes encoding three selected myelin epitopes, which, cloned in *L. lactis* cells under the lactococcal *ptcB* promoter, were demonstrated to efficiently induce oral tolerance by decreasing the clinical symptoms of EAE in rats. Moreover, the conducted animal studies deliver valuable information concerning the optimal administration scheme for *L. lactis*-produced myelin antigens. Further studies on optimizing the dose and feeding regimen as well as peptide expression and delivery will be needed to determine whether increase of the therapeutic effect of *L. lactis* preparations could be obtained. Nonetheless, application of *Lactococcus* bacteria, which have a documented safe effect on human and animal health, for production of mammalian myelin epitopes and oral administration to induce immunotolerance seems to be an attractive means of treatment of autoimmune diseases. Data obtained in this work creates further perspectives for seeking new therapies for multiple sclerosis via the oral tolerance phenomenon.
The authors declare that they have no conflict of interests.

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