

| Applied and Industrial Microbiology | Announcement

Whole-genome sequencing and characterization of human fecal isolate *Lacticaseibacillus casei* **LC130 from NORDBIOTIC collection**

Agnieszka K. Szczepankowska,[1](#page-1-0) Bożena Cukrowska,[2](#page-1-0) Tamara Aleksandrzak-Piekarczyk[1](#page-1-0)

AUTHOR AFFILIATIONS See affiliation list on p. [2.](#page-1-0)

ABSTRACT We report the complete genome sequence of *Lacticaseibacillus casei* LC130, isolated from a healthy human fecal sample and part of the NORDBIOTIC collection. The 2.969 Mb genome of LC130 includes genes potentially involved in lactose metabolism and the production of bacteriocins, peptidases, and polyamines, suggesting potential health benefits.

KEYWORDS probiotics, prohealth effects, bacteriocins, gluten-degrading peptidases, polyamines, lactose metabolism, lactic acid bacteria, fecal human isolate

L acticaseibacillus casei LC130, a lactic acid bacterium, was isolated from a healthy

L human fecal sample and included in the NORDBIOTIC collection, deposited in the *acticaseibacillus casei* LC130, a lactic acid bacterium, was isolated from a healthy Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures GmbH (DSM33796). LC130 alleviates symptoms in patients with irritable bowel syndrome (IBS) [\(1\)](#page-1-0) and acute respiratory tract infections [\(2\)](#page-1-0), and reduces immunoreactivity of glutenderived peptides [\(3\)](#page-1-0).

The strain, provided by Nordic Biotic Ltd. (Warsaw, Poland) as part of the NORD-BIOTIC collection, was cultured at 37°C in De Man, Rogosa and Sharpe medium (Oxoid) under aerobic conditions. DNA was extracted using the cetyltrimethylammonium bromide/lysozyme method [\(4\)](#page-2-0). Bacterial cell walls were digested with lysozyme (20 mg/mL; Sigma-Aldrich) and mutanolysin (5 U/mL; A&A Biotechnology) (15 min, 37°C). DNA concentration was measured using a Qubit fluorimeter (Thermo Fisher Scientific) and used for both Illumina and Nanopore sequencing.

Illumina libraries were constructed with the NEB Ultra II FS kit (New England Biolabs) and sequenced in paired-end mode (v.3, 600 cycle chemistry kit) on a MiSeq equipment (Illumina). Quality was checked using FASTQC (v.0.12.0) [\(5\)](#page-2-0), and short reads were filtered using fastp (v.0.23.2) [\(6\)](#page-2-0).

For long-read sequencing, DNA was sheared with a 26G needle, and fragments shorter than 10 kb were excluded using the Short Read Eliminator kit (Circulomics). A 1D library was constructed with the SQK-LSK109 native barcoding expansion kit (EXP-NBD103) and sequenced using an R9.4.1 flowcell on a GridION sequencer (Oxford Nanopore Technologies). Raw Oxford Nanopore Technology (ONT) reads were basecalled using Guppy (v.6.1.3) [\(https://github.com/nanoporetech/pyguppyclien\)](https://github.com/nanoporetech/pyguppyclien) in super accuracy mode. Short (<1 kb) and low quality (average quality score < 12) reads were removed by NanoFilt (v.2.8.0) [\(7\)](#page-2-0) and adapters were trimmed with Porechop (v.0.2.4) [\(https://github.com/rrwick/Porechop\)](https://github.com/rrwick/Porechop). Data quality check was done with NanoPlot (v.1.41.6) [\(7\)](#page-2-0). ONT long-reads were assembled with Trycycler (v.0.5.3) [\(8\)](#page-2-0) and Flye (v.2.9) [\(9\)](#page-2-0), Unicycler (v.0.4.8) [\(10\)](#page-2-0), Raven (v.1.8.1) [\(11\)](#page-2-0) and Miniasm (v.0.3-r179) [\(12\)](#page-2-0), and polished using Racon (v.1.5.0) [\(13\)](#page-2-0) and Medaka (v.1.7.2) [\(14\)](#page-2-0). Circularization and contig rotation were attained by Trycycler (v.0.5.3) and verified by Bandage (v.0.8.1) [\(15\)](#page-2-0) and Tablet

Editor Kenneth M. Stedman, Portland State University, Portland, Oregon, USA

Address correspondence to Tamara Aleksandrzak-Piekarczyk, tamara@ibb.waw.pl.

The authors declare no conflict of interest.

Received 13 May 2024 **Accepted** 13 June 2024 **Published** 2 July 2024

Copyright © 2024 Szczepankowska et al. This is an open-access article distributed under the terms of [the Creative Commons Attribution 4.0 International](https://creativecommons.org/licenses/by/4.0/) license.

(v.1.17.08.17) [\(16\)](#page-2-0). Default parameters were used for all software. ONT and Illumina assemblies were combined and polished using Polypolish (v.0.5.0) [\(17\)](#page-2-0) and POLCA (v.4.0.5) [\(18\)](#page-2-0). The final coverage for the chromosome was 224x and for plasmid 1373x. Illumina and ONT sequencing statistics are presented in Table 1.

The genome revealed a circular chromosome (2,940,740 bp, 47.9% GC) and a circular plasmid (28,032 bp, 44.08% guanine-cytosine), with 2,898 predicted coding sequences, 59 tRNAs, 15 rRNAs, and 3 ncRNAs. Annotation using the NCBI Prokaryotic Genome Annotation Pipeline (v.6.6) [\(19\)](#page-2-0) identified genes encoding enzyme IIA and enzyme IICB components of the lactose-specific phosphotransferase system, a phospho-β-galactosidase, and the complete tagatose pathway. Strong acidifying properties may enhance calcium/iron solubility and absorption by intestinal cells, increasing bone calcium content and alleviating anemia [\(20–22\)](#page-2-0). Four potential class IId bacteriocins and a gene encoding ornithine decarboxylase, converting ornithine to putrescine (a beneficial polyamine), were found, which may tighten intestinal cell junctions [\(23–26\)](#page-2-0), contributing to IBS symptom relief (1). Genes encoding peptidases that could degrade immunogenic gluten-derived peptides were also present (3).

AUTHOR AFFILIATIONS

¹Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw, Poland ²Department of Pathomorphology, The Children Memorial Health Institute, Warsaw, Poland

AUTHOR ORCIDs

Agnieszka K. Szczepankowska b http://orcid.org/0000-0002-8733-3283 Tamara Aleksandrzak-Piekarczyk **b** http://orcid.org/0000-0002-4725-760X

AUTHOR CONTRIBUTIONS

Agnieszka K. Szczepankowska, Conceptualization, Investigation, Visualization, Writing – original draft, Writing – review and editing | Bożena Cukrowska, Validation, Writing – review and editing | Tamara Aleksandrzak-Piekarczyk, Conceptualization, Investigation, Project administration, Supervision, Validation, Writing – review and editing

DATA AVAILABILITY

Complete sequencing data has been deposited in GenBank under BioProject: [PRJNA1059784;](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA1059784) BioSample: [SAMN39212590.](https://www.ncbi.nlm.nih.gov/biosample/SAMN39212590) Whole genomic data is available under accession numbers [CP143264](https://www.ncbi.nlm.nih.gov/nuccore/CP143264.1) (chromosome) and [CP143265](https://www.ncbi.nlm.nih.gov/nuccore/2706394012,2706396635) (pLC130_p1). lllumina SRA reads are available under accession number [SRR27494201.](https://trace.ncbi.nlm.nih.gov/Traces/?view=run_browser&acc=SRR27494201&display=metadata) Oxford Nanopore SRA reads are available under accession number [SRR27494202.](https://trace.ncbi.nlm.nih.gov/Traces/?view=run_browser&acc=SRR27494202&display=metadata)

REFERENCES

- 1. Skrzydło-Radomańska B, Prozorow-Król B, Cichoż-Lach H, Majsiak E, Bierła JB, Kanarek E, Sowińska A, Cukrowska B. 2021. The effectiveness and safety of multi-strain probiotic preparation in patients with diarrhea-predominant irritable bowel syndrome: a randomized controlled study. Nutrients 13:756.<https://doi.org/10.3390/nu13030756>
- 2. Kolesnyk PO, Paliy IH, Sydorchuk LP, Hoda ZP, Ivanchenko NO, Lych OS, Huley NR, Matsyura OI, Slyuzar ZL, Gerasymov SV. 2024. The role of

nutritional support with probiotics in outpatients with symptomatic acute respiratory tract infections: a multicenter, randomized, double[blind, placebo-controlled dietary study. BMC Nutr](https://doi.org/10.1186/s40795-023-00816-8) 10:4. https://doi.org/ 10.1186/s40795-023-00816-8

3. Leszczyńska J, Szczepankowska AK, Majak I, Mańkowska D, Smolińska B, Ścieszka S, Diowksz A, Cukrowska B, Aleksandrzak-Piekarczyk T. 2024. Reducing immunoreactivity of gluten peptides by probiotic lactic acid bacteria for dietary management of gluten-related diseases. Nutrients 16:976.<https://doi.org/10.3390/nu16070976>

- 4. Wilson K. 2001. Preparation of genomic DNA from bacteria. Curr Protoc Mol Biol [Chapter 2:Unit. https://doi.org/10.1002/0471142727.](https://doi.org/10.1002/0471142727.mb0204s56) mb0204s56
- 5. Andrews S. 2010. FastQC: a quality control tool for high throughput [sequence data. Available from: http://www.bioinformatics.babraham.ac.](http://www.bioinformatics.babraham.ac.uk/projects/fastqc/) uk/projects/fastqc/
- 6. Chen S, Zhou Y, Chen Y, Gu J. 2018. Fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics [34:i884–i890. https://doi.org/10.1093/](https://doi.org/10.1093/bioinformatics/bty560) bioinformatics/bty560
- 7. De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics [34:2666–2669. https://doi.org/10.1093/bioinformatics/](https://doi.org/10.1093/bioinformatics/bty149) bty149
- 8. Wick RR, Judd LM, Cerdeira LT, Hawkey J, Méric G, Vezina B, Wyres KL, Holt KE. 2021. Trycycler: cnsensus long-read assemblies for bacterial genomes. Genome Biol [22:266. https://doi.org/10.1186/s13059-021-](https://doi.org/10.1186/s13059-021-02483-z) 02483-z
- 9. Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error[prone reads using repeat graphs. Nat Biotechnol](https://doi.org/10.1038/s41587-019-0072-8) 37:540–546. https://doi. org/10.1038/s41587-019-0072-8
- 10. Wick R.R, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595.<https://doi.org/10.1371/journal.pcbi.1005595>
- 11. Vaser R, Šikić M. 2021. Time- and memory-efficient genome assembly with raven. Nat Comput Sci [1:332–336. https://doi.org/10.1038/s43588-](https://doi.org/10.1038/s43588-021-00073-4) 021-00073-4
- 12. Li H. 2016. Minimap and miniasm: fast mapping and *de novo* assembly [for noisy long sequences. Bioinformatics](https://doi.org/10.1093/bioinformatics/btw152) 32:2103–2110. https://doi.org/ 10.1093/bioinformatics/btw152
- 13. Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate *de novo* genome assembly from long uncorrected reads. Genome Res 27:737– 746.<https://doi.org/10.1101/gr.214270.116>
- 14. Wright C, Wykes M. 2023. Medaka: sequence correction provided by ONT research. [https://github.com/nanoporetech/medaka.](https://github.com/nanoporetech/medaka)
- 15. Wick RR, Schultz MB, Zobel J, Holt KE. 2015. Bandage: interactive visualization of *de novo* genome assemblies. Bioinformatics 31:3350– 3352.<https://doi.org/10.1093/bioinformatics/btv383>
- 16. Milne I, Bayer M, Cardle L, Shaw P, Stephen G, Wright F, Marshall D. 2010. Tablet--next generation sequence assembly visualization. Bioinformatics 26:401–402.<https://doi.org/10.1093/bioinformatics/btp666>
- 17. Wick RR, Holt KE. 2022. Polypolish: short-read polishing of long-read [bacterial genome assemblies. PLoS Comput Biol](https://doi.org/10.1371/journal.pcbi.1009802) 18:e1009802. https:// doi.org/10.1371/journal.pcbi.1009802
- 18. Zimin AV, Puiu D, Luo M-C, Zhu T, Koren S, Marçais G, Yorke JA, Dvořák J, Salzberg SL. 2017. Hybrid assembly of the large and highly repetitive genome of *Aegilops tauschii*, a progenitor of bread wheat, with the [MaSuRCA mega-reads algorithm. Genome Res](https://doi.org/10.1101/gr.213405.116) 27:787–792. https://doi. org/10.1101/gr.213405.116
- 19. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614– 6624.<https://doi.org/10.1093/nar/gkw569>
- 20. Pérez-Conesa D, López G, Ros G. 2007. Effect of probiotic, prebiotic and synbiotic follow-up infant formulas on iron bioavailability in rats. Food Sci Technol Int 13:69–77.<https://doi.org/10.1177/1082013207075465>
- 21. Pérez AV, Picotto G, Carpentieri AR, Rivoira MA, Peralta López ME, Tolosa de Talamoni NG. 2008. Minireview on regulation of intestinal calcium absorption. Emphasis on molecular mechanisms of transcellular pathway. Digestion 77:22–34.<https://doi.org/10.1159/000116623>
- 22. Scholz-Ahrens KE, Ade P, Marten B, Weber P, Timm W, Açil Y, Glüer C-C, Schrezenmeir J. 2007. Prebiotics, probiotics, and synbiotics affect mineral absorption, bone mineral content, and bone structure. J Nutr 137:838S–46S.<https://doi.org/10.1093/jn/137.3.838S>
- 23. Mikelsaar M, Stsepetova J, Mikelsaar R-H, Truusalu K, Smidt I, Hütt P, Rätsep M, Songisepp E. 2020. Polyamines of human strain *Lactobacillus plantarum* inducia induce modulation of innate immune markers. J Funct Foods72:104064.<https://doi.org/10.1016/j.jff.2020.104064>
- 24. Heeney DD, Zhai Z, Bendiks Z, Barouei J, Martinic A, Slupsky C, Marco ML. 2019. *Lactobacillus plantarum* bacteriocin is associated with intestinal and systemic improvements in diet-induced obese mice and maintains epithelial barrier integrity *in vitro*. Gut Microbes 10:382–397. <https://doi.org/10.1080/19490976.2018.1534513>
- 25. Matsumoto M, Kurihara S, Kibe R, Ashida H, Benno Y. 2011. Longevity in mice is promoted by probiotic-induced suppression of colonic senescence dependent on upregulation of gut bacterial polyamine production. PLoS ONE [6:e23652. https://doi.org/10.1371/journal.pone.](https://doi.org/10.1371/journal.pone.0023652) 0023652
- 26. Gu Q, Yan J, Lou Y, Zhang Z, Li Y, Zhu Z, Liu M, Wu D, Liang Y, Pu J, Zhao X, Xiao H, Li P. 2024. Bacteriocins: curial guardians of gastrointestinal [tract. Compr Rev Food Sci Food Saf](https://doi.org/10.1111/1541-4337.13292) 23:e13292. https://doi.org/10.1111/ 1541-4337.13292