The role of gut microbiota metabolites in the regeneration and protection of nervous tissue: a narrative review

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Abstract

The gut microbiota modulates various physiological functions in the human body, including digestion, immune regulation, gut barrier maintenance, and even nervous system activity. The bidirectional communication between gut microbes and the brain, known as the microbiota–gut–brain axis, is crucial for balanced metabolism. Recent studies have indicated that gut microbiota metabolites, such as short-chain fatty acids, indole derivatives, neurotransmitters, and other bioactive compounds, can positively impact neurogenesis, myelination, and axonal regeneration, suggesting their potential in therapeutic strategies for neuroprotection and neuroregeneration. Despite the growing number of studies on gut microbiota metabolites, understanding their role in neuroprotective mechanisms remains limited. This article reviews the classification, production, functions and therapeutic potential of the most well-known gut microbiota metabolites, as well as their impact on neurogenesis, synaptogenesis, energy metabolism, immune modulation, and blood–brain barrier integrity, which will provide a foundation for the study of gut microbiota metabolites in the field of biomedical engineering.

Key words: biomedical engineering; gut microbiota; metabolites; microbiota; neuroprotection; neuroregeneration; regenerative medicine; regenerative therapy

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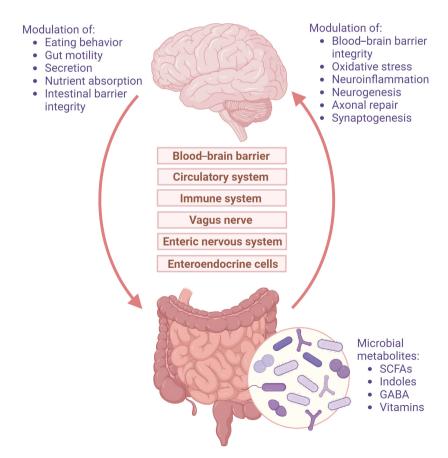
INTRODUCTION

The human body contains, on average, more microbial cells than human cells do, with approximately 38 trillion bacteria compared with 30 trillion human cells.¹ Additionally, the total genome size of human commensal microbes far exceeds that of the host genome. These diverse microbial communities, collectively known as the microbiota, include not only bacteria but also archaea, eukaryotes, and viruses. Microorganisms inhabit various organs of the human body, such as the distal tracts of the genitourinary system, nasal cavities, and skin, but the majority reside in the gastrointestinal tract (GIT).² The gut microbiota shares a mutually beneficial relationship with its host, contributing to tissue homeostasis through numerous physiological functions. These functions include digestion, nutrient absorption, vitamin synthesis, maintenance of intestinal barrier integrity, protection against pathogens, cholesterol metabolism and modulation of both the immune system and the central nervous system (CNS).³ The broad genetic and metabolic potential of the gut microbiota supports its involvement in nearly every aspect of human biology, from health maintenance and development to regeneration, aging, and disease.⁴

The roles of the gut microbiota extend far beyond the boundaries of the GIT. Intestinal microbes continuously interact with the gut and distant organs of the host through metabolites and signaling molecules produced by both the host and the gut microbes.⁵ Growing evidence has revealed the bidirectional communication between the gut microbiome and the CNS, known as the "microbiota-gut-brain axis".⁵⁻⁷ Communication between the gut microbiota and the brain occurs through various pathways, including the neuroendocrine system, vagus nerve, enteric nervous system (ENS), immune system, and circulatory system, via the production of neuroactive substances, metabolites, and hormones (Figure 1). Studies have demonstrated that the gut microbiota can produce or stimulate the production of neurotransmitters, including serotonin, dopamine, acetylcholine, and

y-aminobutyric acid (GABA), as well as neuroactive metabolites such as short-chain fatty acids (SCFAs) and vitamins.⁸⁻¹¹ These substances influence both enteric neurons in the gut and neurons in the CNS. Overall, the interaction between the gut microbiota and the CNS is modulated directly by immune cells and nerve fibers and indirectly through the production of metabolites and hormones that can bypass the blood-brain barrier (BBB). Gut microbes and their metabolites interact with immune cells within gut-associated lymphoid tissues, thus regulating the production of proinflammatory and anti-inflammatory mediators.¹² The gut microbiota also indirectly impacts non-neuronal cells in the CNS, such as microglia, astrocytes, oligodendrocytes, and oligodendrocyte precursor cells.¹³⁻¹⁵ These cells are essential for maintaining brain health and function through various roles, such as immune regulation, synaptic pruning, myelination, and neuronal support. When influenced by microbial metabolites or immune signals originating from the gut, these CNS cells can undergo changes in their activity levels, cytokine

production, or responsiveness to signaling molecules.⁴ Such interactions highlight the broader impact of microbiota metabolites in microbiota-gut-brain communications beyond neurons alone, contributing significantly to the overall balance and functionality of the CNS. Some gut microbiota metabolites have emerged as significant players in the modulation of BBB integrity, particularly within the context of neuroprotection and neuroregeneration.¹⁶ By interacting with endothelial cells that constitute the BBB. microbial metabolites (e.g., SCFAs) can regulate the expression of tight junction (TJ) proteins, thereby enhancing barrier function.¹⁷ This regulation is crucial, as a compromised BBB can lead to increased permeability and neuroinflammation, which are implicated in neurodegenerative diseases and neurological injuries. The potential of gut microbiota metabolites to enhance BBB integrity suggests therapeutic benefits for promoting neuroprotection. This involves limiting the entry of harmful substances into the brain and supporting neuroregeneration processes.



Microbiota-gut-brain axis

Figure 1: The microbiota-gut-brain axis

Note: Created with BioRender.com. GABA: γ-Aminobutyric acid; SCFAs: short-chain fatty acids.

Several studies have highlighted the significant role of gut microbiota metabolites as neuroprotective agents that promote nerve regeneration and repair.^{18, 19} However, despite these findings, there has been no comprehensive summary of the functions of gut microbiota metabolites related to the protection and regeneration of nervous tissue. This gap has limited our understanding of the involvement of the gut microbiota in neuroregeneration. In this review, we aim to systematize the knowledge of gut microbiota metabolites within the microbiota-gut-brain axis. We begin with a brief introduction to how microbial metabolites impact nervous system function, followed by the classification of these metabolites. Next, we discuss their functions in detail, with a particular focus on their roles in regulating neurogenesis, synaptogenesis, and energy metabolism; modulating the local and systemic immune systems; and influencing the BBB. Finally, we discuss future research directions, therapeutic strategies and potential avenues for further exploration in this field.

SEARCH STRATEGY

To conduct this literature review, various databases containing up-to-date information in the field were consulted: Web of Science, PubMed, ScienceDirect, Cochrane (Wiley), and Scopus were searched, and studies published between 2014 and 2024 were the focus. The search utilized specific keywords: "microbiota-gut-brain axis," "gut microbiota metabolites," "mycobiome," "neuroprotection," "regeneration," "neurotransmitter production," "bloodbrain barrier integrity," "neurogenesis," "axonal repair," and "synaptogenesis." MeSH guidelines were followed to ensure the relevance of the literature reviewed. The search period was limited to articles published between 2014 and 2024, which guarantees the timeliness and relevance of the information included in this review. The following exclusion criteria were employed: (1) studies outside the period analyzed; (2) topics presented outside the scope of the review; and (3) books, conference proceedings, doctoral theses, and abstracts. Articles meeting scientific methodological standards and relevant to the review's subsections were included. Hence, this narrative review provides a summary of the involvement of gut microbiota metabolites in neuroprotection and neuroregeneration.

ROLE OF MICROBIAL METABOLITES IN THE MICROBIOTA-GUT-BRAIN AXIS

The gut microbiota plays an essential role in the

physiological development and maintenance of homeostasis in the peripheral and central nervous systems, which is why any significant disruption of the microbiota composition can lead to various serious disorders. For example, an imbalance in the gut microbiota, known as dysbiosis, has been associated with various neurological and psychological conditions, such as anxiety, depression, autism spectrum disorder and neurodegenerative diseases such as multiple sclerosis, Parkinson's disease and Alzheimer's disease.^{4, 20} These findings underscore the significant role of the gut microbiota in crucial neurodevelopmental processes, including neurogenesis, myelination, microglial maturation, development and maintenance of BBB integrity.⁷ One of the key mechanisms underlying microbiota-gut-brain crosstalk is the production of bioactive metabolites (Figure 2). The primary signaling pathway involving microbial metabolites occurs through the vagus nerve and spinal nerves of the autonomic nervous system. Microbial metabolites can signal directly to the vagus nerve via specialized enteroendocrine cells called "neuropod cells". Neuropod cells act as intermediaries, translating microbial signals into neural impulses that can be directly transmitted to the brain.²¹ Notably, microbiota metabolites also directly influence enteric neurons and glial cells, which are integral components of the ENS, a part of the autonomic nervous system located in the GIT. The ENS plays a crucial role in coordinating gut functions, including motility, digestion, gut barrier function and intestinal mucosal immunity. Two ganglionated plexuses of the ENS, the submucosal plexus (Meissner's plexus) and the myenteric plexus (Auerbach's plexus), comprise an abundance of catecholaminergic, neuropeptideexpressing and GABAergic neurons.²² The activity of these neurons can be modulated by the gut microbiota, which can produce neurotransmitters. Moreover, enteric neurons express Toll-like receptors, free fatty acid receptors and aryl hydrocarbon receptors (AhRs), which respond to other microbial metabolites, such as lipopolysaccharides (LPS), SCFAs and indole derivatives.^{4, 22} Microbial metabolites, including norepinephrine, indole, indole-3-aldehyde, butyric acid, isobutyric acid and isovaleric acid, can also stimulate enterochromaffin cells, which release serotonin (also known as 5-hydroxytryptamine (5-HT)), a neurotransmitter involved in various physiological processes, including mood regulation and gastrointestinal function.⁷

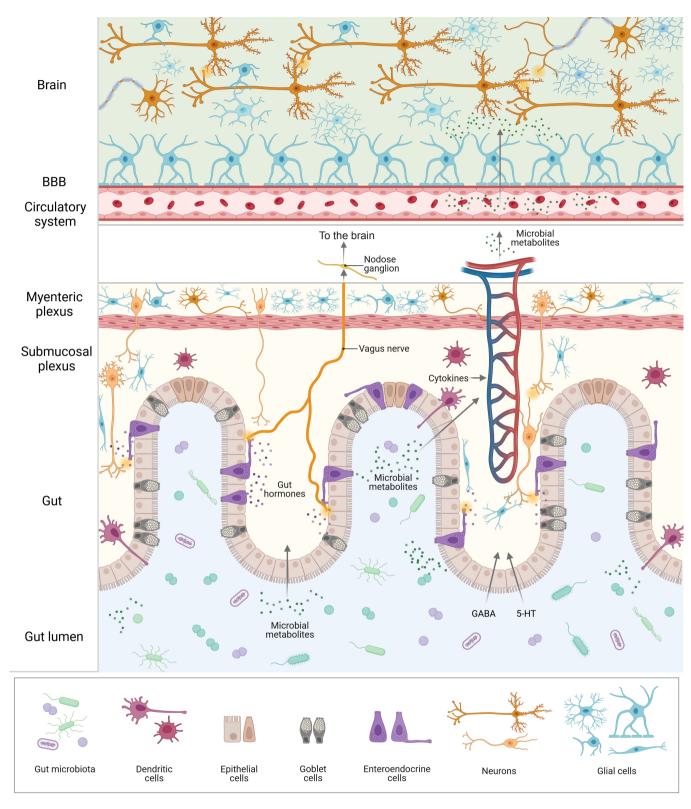


Figure 2: Mechanisms of action of gut microbiota metabolites on the nervous system

Note: Gut microbiota metabolites impact the nervous system through multiple pathways. Directly, microbial metabolites influence enteroendocrine cells in the gastrointestinal tract, which respond by releasing hormones and peptides that affect the enteric nervous system and communicate with the central nervous system (CNS). The vagus nerve transmits signals from microbial metabolites in the gut directly to the brain. Furthermore, dendritic cells in the gut interact with these metabolites, modulating cytokine production and initiating immune responses that can indirectly affect neuroinflammation and neuroimmune signaling. Microbial metabolites can also enter the bloodstream, allowing systemic circulation to transport them to the brain. Upon reaching the brain, they cross the blood–brain barrier (BBB) and affect CNS activity. Microbial metabolites can also affect the permeability of the BBB, a key factor in regulating what substances can enter the brain. This alteration in BBB permeability can indirectly impact brain function by modifying the microenvironment within the CNS. Created with BioRender.com. GABA: γ -Aminobutyric acid; 5-HT: serotonin.

Another possible underlying pathway of microbiotagut-brain communication involves the circulatory system. Some microbial metabolites can cross the intestinal barrier and enter the bloodstream. From there, they can bypass the BBB and influence the CNS. This mechanism illustrates how the gut microbiota can indirectly affect brain function and behavior through the production of bioactive metabolites. For example, SCFAs such as butyric acid have been shown to cross the intestinal barrier and the BBB via monocarboxylate transporters located on endothelial cells, enabling SCFAs to interact with neuronal and glial receptors and influence gene expression in the brain.²³ In addition to bypassing the BBB and the intestinal barrier, microbial metabolites play an important role in maintaining barrier integrity, which is linked to the regulated passage of molecules and nutrients from the bloodstream to the brain. Several studies have highlighted the ability of microbial metabolites, such as SCFAs, branched-chain fatty acids (BCFAs) and indole derivatives, to increase the expression of TJ proteins such as claudins, occludin and zonula occludens-1 (ZO-1), thus regulating intestinal and BBB integrity.^{16, 24-26} However, it is important to note that not all microbial metabolites can directly access the CNS though the bloodstream. Larger molecules, such as exopolysaccharides (EPSs), and neurotransmitters, including serotonin, GABA, acetylcholine, and noradrenaline, cannot cross the BBB, but they can impact enteric neurons and the vagus nerve, thus indirectly modulating CNS function.²⁷ In contrast, derivatives and precursors of neurotransmitters such as tyrosine or tryptophan can bypass the BBB, entering systemic circulation to be utilized for synthesizing neurotransmitters within the CNS.²⁷

Microbiota-derived metabolites can also influence CNS functions by affecting immune responses and inflammation. They can trigger the release of pro- or anti-inflammatory cytokines, activate immune cells that circulate through the bloodstream, and impact BBB function, thus modifying the inflammatory status of brain cells.^{12, 28} Metabolite-induced secretion of cytokines by immune cells can locally influence enteric neurons and modify vagus nerve signaling to the brain.²⁹ Research has shown that intestinal macrophages and their cytokine outputs can alter how enteric neurons respond to inflammatory signals and impact the apoptosis of these neurons. Moreover, cytokines affect the activity of enteric glial cells, which play crucial roles in supporting and regulating the function of enteric neurons by producing glial cell-derived neurotrophic factor. The production of this neurotrophic factor induced by SCFAs has been reported to stimulate the

production of interleukin-22 (IL-22), thereby promoting the protection and restoration of the epithelial barrier.²⁹ Furthermore, microbial metabolites are recognized by Toll-like receptors expressed not only in enteric neurons but also in the smooth muscle cells of the intestinal wall and dendritic cells.²² Within the CNS, the microbial metabolite-induced immune response can influence microglial phenotype and function, impacting their roles in synaptic pruning, neuroinflammation and neuroprotection.⁴ For example, tryptophan metabolites such as indole derivatives can exert neuroprotective effects by modulating the kynurenine pathway, influencing the nuclear factor kappa B (NF-κB) signaling pathway and oxidative stress in the brain.³⁰

Finally, microbial metabolites within the microbiotagut-brain axis can influence energy metabolism. For example, SCFAs serve as key energy sources for intestinal cells. They can also cross the BBB and affect the energy metabolism of neurons and glial cells.¹⁷ SCFAs can enhance mitochondrial function in brain cells, potentially boosting cognitive processes and protecting against neurological disorders. Moreover, SCFAs can act as signaling molecules that can modulate appetiteregulating hormones such as leptin and ghrelin.²³ By binding to specific receptors on enteroendocrine cells in the gut, SCFAs can stimulate the release of peptides that affect hunger and satiety signals sent to the brain. The gut microbiota has been reported to produce menaguinones and group B vitamins, which serve as critical cofactors in a wide array of metabolic reactions.³¹⁻³³

As previously mentioned, the gut microbiota comprises a diverse range of microorganisms. However, the bioactive metabolites produced by archaea and fungi remain relatively understudied. Therefore, this review focuses mostly on bacterial metabolites. Furthermore, our scope is specifically aimed at discussing the role of gut microbiota metabolites in protecting and regenerating nervous tissue, which is why we do not discuss microbial metabolites that exhibit detrimental effects on nervous system function. The gut microbiota comprises six primary groups of bacteria: Bacillota (previously known as Firmicutes), Bacteroidota (previously known as Bacteroidetes), Actinomycetota (previously known as Actinobacteria), Pseudomonadota (previously known as Proteobacteria), Fusobacteriota (previously known as Fusobacteria), and Verrucomicrobiota (previously known as Verrucomicrobia), with Bacillota and Bacteroidota being the predominant phyla.^{34, 35} Within these groups, bacteria from families such as Lachnospiraceae, Oscillospiraceae (previously known as Ruminococcaceae), Bacteroidaceae, Prevotellaceae, and genera such as Faecalibacterium, Bacteroides, Bifidobacterium, Lactobacillus, Prevotella, Roseburia, and Akkermansia have been reported to produce healthpromoting metabolites.^{7, 36} The fungal community, known as the gut mycobiome, represents a smaller yet significant portion of the gut microbiota.³⁷ The most prevalent fungal phylum in the gut is Ascomycota, followed by Zygomycota and Basidiomycota. Within these groups, the genera Candida, Saccharomyces, Malassezia, Cryptococcus, Aspergillus, Galactomyces, Trichosporon, and Cladosporium are commonly found in the GIT.^{38, 39} Gut fungi hold significant potential in the production of a wide array of metabolites that can be highly beneficial for both human health and gut bacteria. The gut mycobiota is known to produce various bioactive compounds, including β -1,3-glucan, a polysaccharide with powerful immunomodulatory properties. They also play crucial roles in the synthesis of vitamins, farnesol, tyrosol, and fatty acids that can support the growth and viability of beneficial gut bacteria.^{37, 38} The archaea in the human gut microbiota are methane producers, with the predominant groups being Methanobacteriales, such as Methanobrevibacter smithii and Methanosphaera stadtmanae, and Methanomassiliicoccales, such as members of the Methanomethylophilaceae family. Recent studies have shown that not all archaea in the human gut are methanogens, with viable haloarchaeal strains from the genus *Haloferax* being isolated from human feces.⁴⁰ It has been suggested that gut archaea, similar to gut bacteria, may produce beneficial metabolites such as SCFAs, tryptophan derivatives, GABA, and deconjugated bile acids.⁴¹ However, research in this area is still limited and warrants further exploration.

Overall, microbial metabolites with healthpromoting potential within the microbiota-gut-brain axis can be categorized into several major groups. Specifically, fatty acids (such as SCFAs, BCFAs, and conjugated polyunsaturated fatty acids), lactic acid, vitamins (including vitamins B and menaquinones), tryptophan derivatives (such as indole derivatives and 3-hydroxyanthranilic acid (3-HAA)), neurotransmitters (GABA, dopamine, and norepinephrine), deconjugated bile acids, polysaccharides (bacterial and fungal EPSs, such as dextran and β -glucans), fungal-derived secondary metabolites (tyrosol and farnesol) and polyphenol-derived secondary metabolites (equol and urolithins) were described. These metabolites influence nervous system function through a variety of mechanisms, ranging from affecting the maturation

of microglia and neurogenesis to modulating synaptic plasticity and neurotransmitter synthesis. A summary of health-promoting microbial metabolites, including the taxa (families, genera or species) of microbes capable of producing these compounds, along with their effects on neurological function and the specific mechanisms involved, is provided in **Table 1.**

NEUROPROTECTIVE MECHANISMS INDUCED BY GUT MICROBIOTA METABOLITES

Neuroprotection is considered one of the key therapeutic strategies for slowing neurological disease progression and promoting regeneration processes. The underlying mechanisms of neuroprotection involve mitigating oxidative stress, reducing neuroinflammation and mitochondrial dysfunction, improving BBB function, preventing glutamate excitotoxicity, correcting protein misfolding, maintaining proper autophagy, and inhibiting apoptosis.⁴² Various microbial metabolites have been reported to influence these neuroprotective pathways in multiple beneficial ways.⁴³⁻⁴⁷ For example, SCFAs, such as acetic, propionic, butyric and valeric acids, as well as BCFAs, such as isobutyric and isovaleric acids, have demonstrated significant anti-inflammatory and antioxidant properties.^{7, 17, 48} Butyrate, in particular, can inhibit histone deacetylases, leading to the upregulation of genes involved in antioxidant defense, thus reducing oxidative stress. The influence of SCFAs on epigenetic regulation through histone acetylation and DNA methylation modulates the NF-κB signaling pathway and inflammatory cytokine production, reducing neuroinflammation.⁴⁹ An in vitro characterization of gut microbiota-derived bacterial strains revealed that Parabacteroides distasonis MRx0005 and Megasphaera massiliensis MRx0029 can produce butyric and valeric acids, which decrease neuroinflammation in glioblastoma cells and oxidative stress in neuron-like cells. Butyric acid specifically decreases IL-6 secretion in the presence of LPS.¹⁸ Furthermore, microbiotaderived butyrate modulates mitochondrial activity in a lymphoblastoid cell line model of autism spectrum disorder, rescuing energy metabolism during oxidative stress.⁵⁰ Additionally, butyrate was found to promote the expression of TJ proteins, such as occludin and ZO-1, which are essential for the integrity of the BBB and intestinal barrier.48, 51 In a mouse model of traumatic brain injury, the positive effect of butyrate on TJ proteins and BBB permeability resulted in the attenuation of neurological deficits, brain edema, and neuronal damage.²⁴

Table 1: Microbial metabolites, the gut microbiota, and their health-promoting potential within the microbiota-gut-brain axis

Gut metabolites	Gut microbiota	Effects	Mechanism of action	Reference
Short-chain fatty acids (SCFAs):	Acetobacteraceae	个Neurogenesis	↑Proliferation of neural progenitor cells	13, 17, 18,
Acetic acid	Lachnospiraceae	↓ Neuroinflammation	↑BDNF expression	24, 37, 43,
 Propionic acid 	Oscillospiraceae	↓Neuronal apoptosis	↑Neurotransmitter synthesis	48-50, 52,
 Butyric acid 	Erysilotrichaceae	↓Oxidative stress	个GAP-43	53, 105, 106,
	Roseburia	↑Mitochondrial biogenesis	\downarrow Activity of histone deacetylase	108, 113, 115
	Bacteroides	↑Dorsal root ganglia outgrowth	↓ Proinflammatory cytokines	
	Bifidobacterium	↑Integrity of BBB and intestinal barrier	Cholecystokinin-dependent stimulation of	
	Blautia	↓ Demyelination	the vagus nerve	
	Eubacterium	↑Remyelination Anti-inflammatory effects	↓Activation of microglia and astrocytes ↑Maturation of microglia	
	Clostridium butyricum Faecalibacterium prausnitzii	Immunomodulation	↑Maturation of microglia	
	Parabacteroides distasonis	Modulation of enteroendocrine cells	↑ Survival of Schwann cells	
	Akkermansia muciniphila	Modulation of synaptic plasticity and cognitive	↑TJ proteins	
	Saccharomyces, Aspergillus	functions	Energy source for cells	
		Modulation of neuronal energy metabolism	Agonists of FFARs	
Branched-chain fatty acids	Lachnospiraceae	↓ Neuroinflammation	↓ Activation of microglia and astrocytes	17, 51, 118,
(BCFAs):	Oscillospiraceae	↓Neuronal apoptosis	↓ Pro-inflammatory cytokines	128
 Isobutyric acid 	Peptostreptococcaceae	↓Oxidative stress	\downarrow Activity of histone deacetylase	
 Isovaleric acid 	Bacteroidaceae	↑Integrity of BBB and intestinal barrier	↑Neurotransmitter synthesis	
	Blautia	Anti-inflammatory effects	个TJ proteins	
	Ruminococcus	Modulation of enteroendocrine cells	Energy source for cells	
	Bacteroides	Modulation of neuronal energy metabolism	Agonists of FFARs	
	Lactobacillaceae	↓Neuroinflammation	↓ Proinflammatory cytokines	55, 56, 109-
	Bifidobacteriaceae	↑Neurogenesis	Veutrophil infiltration	111, 129-131
	Streptococcaceae	↑Axonal regeneration	↑SIRT-mediated BDNF expression	
	Enterococcaceae	↑Neuroplasticity and cognitive functions	↑GABA expression	
		↑Intestinal barrier integrity	↓ Glutamate excitotoxicity	
		Immunomodulation Modulation of neuronal energy metabolism	Stimulation of the vagus nerve	
Conjugated polyunsaturated fatty	Lactinlantibacillus plantarum	↓ Neuroinflammation	↑Proliferation of neural progenitor cells	57-61
	Limosilactobacillus reuteri	↓Oxidative stress	↑Anti-inflammatory cytokines	57 01
	Lactococcus lactis	↓ Lipid peroxidation	↓ Astrogliosis	
 Conjugated linolenic acid (CLNA) 		Anti-inflammatory effects	↓ ROS production	
		Anti-aggregative effect against $A\beta_{1-42}$	↑PPARα-induced peroxisomal β-oxidation	
Group B vitamins:	Bacteroides fragilis	↓ Neuroinflammation	↑Neurotransmitter synthesis	31, 33, 62-
• Thiamin (B1)	Segatella copri	↑Neuronal survival	↓ROS production	67, 112
• Riboflavin (B2)	Ruminococcus lactaris	↓ Demyelination	\downarrow Glutamate excitotoxicity	
• Niacin (B3)	Corynebacterium glutamicum	↓Neuronal death	ightarrowNF-κB-mediated transcription of	
	Propionibacterium freudenreichi	↓Oxidative stress	inflammatory mediators	
	Faecalibacterium prausnitzii	Cytoprotective and anti-inflammatory effects	个TJ proteins	
	Lactobacillus acidophilus	↑Integrity of BBB and intestinal barrier	Cofactors of various enzymatic reactions in	
()	Lactobacillus delbrueckii	Modulation of neuronal energy metabolism	the metabolism of carbohydrates, fats, and	
• Cobalamin (B12)	Lactiplantibacillus plantarum		amino acids	
	Streptococcus thermophilus		Regulation of tryptophan metabolism via the	
	Lactococcus lactis	Neuroinflommation	kynurenine pathway	22 44 68
	Prevotella Bronionibactorium fraudanraichi	↓Neuroinflammation ↓Neuronal apoptosis		32, 44, 68, 69, 114
(vitamin K2)	Propionibacterium freudenreichi Bacteroides fragilis	\oint Oxidative stress	↓ROS production	09, 114
	Limosilactobacillus reuteri	Anti-inflammatory effects	↓iNOS expression	
	Lactococcus lactis	Mitochondrial protection	↑ SOD and GSH-Px activity	
	Leuconostoc lactis	↑Integrity of BBB and intestinal barrier	↑Endothelial progenitor cell proliferation	
	Bacillus subtilis	Anti-aggregative effect against $A\beta_{1,42}$ and	and migration	
		α-synuclein	↑Sphingolipid synthesis	
3-Hydroxyanthranilic acid (3-HAA)	Pseudomonas	↓Aging	↑Nrf2/SKN-1 oxidative stress response	28, 30, 76,
	Burkholderia	↑ Lifespan	pathway	77, 132
	Stenotrophomonas	↑Resistance to oxidative stress	↓PI3K/NF-кB signaling pathways	
	Xanthomonas	Immunomodulation		
	Shewanella			
	Bacillus			
		Modulation of gastrointestinal motility	ENS excitatory neurotransmitter	9, 46, 119-
γ-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum	Modulation of synaptic transmission in the ENS	CNS inhibitory neurotransmitter	9, 46, 119- 122, 133, 134
γ-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum Bifidobacterium adolescentis	Modulation of synaptic transmission in the ENS Regulation of anxiety and depression	CNS inhibitory neurotransmitter ↓Neuron excitability	
γ-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum Bifidobacterium adolescentis Lactiplantibacillus plantarum	Modulation of synaptic transmission in the ENS Regulation of anxiety and depression Anti-nociceptive properties	CNS inhibitory neurotransmitter	
γ-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum Bifidobacterium adolescentis Lactiplantibacillus plantarum Levilactobacillus brevis	Modulation of synaptic transmission in the ENS Regulation of anxiety and depression	CNS inhibitory neurotransmitter ↓Neuron excitability	
y-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum Bifidobacterium adolescentis Lactiplantibacillus plantarum Levilactobacillus brevis Streptococcus thermophilus	Modulation of synaptic transmission in the ENS Regulation of anxiety and depression Anti-nociceptive properties	CNS inhibitory neurotransmitter ↓Neuron excitability	
γ-Aminobutyric acid (GABA)	Bacillus Bacteroides fragilis Bifidobacterium angulatum Bifidobacterium adolescentis Lactiplantibacillus plantarum Levilactobacillus brevis	Modulation of synaptic transmission in the ENS Regulation of anxiety and depression Anti-nociceptive properties	CNS inhibitory neurotransmitter ↓Neuron excitability	

Table 1 Continued

Gut metabolites	Gut microbiota	Effects	Mechanism of action	Reference
Indole derivatives:	Lactobacillus	个Neurogenesis	↑BDNF expression	8, 19, 25, 26
Indole-3-aldehyde	Bifidobacterium	↑Axonal regeneration	个NGF expression	45, 70-75,
Indole-3-acetaldehyde	Bacteroides ovatus	↑Neurite outgrowth	↑Neurotransmitter synthesis	107, 116-11
Indole-3-acetic acid	Bacteroides fragilis	↑Functional integration of neurons in the	$ m \uparrow Neutrophil$ chemotaxis toward the dorsal	
Indole-3-lactic acid	Clostridium sporogenes	hippocampus	root ganglia	
 Indole-3-propionic acid 	Clostridium caloritolerans	↑Sensory neurological recovery	↑Synaptic plasticity	
Indoleacrylic acid	Peptostreptococcus anaerobius	↑Spatial memory	$ m \uparrow$ Inhibitory synaptic transmission in the	
• Kynurenic acid	Peptostreptococcus russellii	↑Social behavior	hippocampus	
 5-hydroxytryptophan (5-HTP) Tryptamine 		↑Integrity of BBB and intestinal barrier	个ERK1 signaling	
		\downarrow Neuroinflammation	个AhR signaling	
		\downarrow Neuronal apoptosis	个TJ proteins	
		\downarrow Oxidative stress	\uparrow PGC-1 α -mediated mitochondrial	
		Mitochondrial protection	biogenesis	
		Modulation of enteroendocrine cells	↓Synaptic overpruning	
			\downarrow ROS production	
			\downarrow Endothelial dysfunction	
			↓Activation of microglia	
			ightarrowNF-кB-mediated transcription of	
			inflammatory mediators	
			个GPR30/AMPK/SIRT1 pathway	
			Tryptophan metabolism via the kynurenine	
			pathway	
Catecholamines:	Lactiplantibacillus plantarum	Modulation of synaptic plasticity and cognitive	Neurotransmitters	119, 123,
• Dopamine	Bifidobacterium longum	functions		135, 136
Norepinephrine	Bacillus subtilis	Regulation of intestinal motility, secretion and		
	Escherichia coli	enteric neurotransmission		
	Serratia marcescens			
	Streptococcus thermophilus			
	Enterococcus faecium			
	Eggerthella lenta			
	Proteus vulgaris			
Deconjugated bile acids	Lactobacillaceae	\downarrow Oxidative stress	\downarrow ROS production	79-83
	Bifidobacteriaceae	Modulation of BBB and intestinal barrier	\downarrow Apoptosis of endothelial cells	
	Streptococcaceae	Modulation of enteroendocrine cells	FXR-mediated inhibition of proinflammatory	
	Oscillospiraceae	Regulation of cholesterol absorption	cytokine expression	
	Lactococcus lactis	Immunomodulation		
Exopolysaccharides (EPSs)	Lactobacillus	Antioxidant and immunomodulatory effects	\downarrow ROS production	38, 84-86,
	Bifidobacterium	Anti-aggregative effect against $A\beta_{1-42}$	\downarrow iNOS expression	88-90, 137
	Streptococcus	Mitochondrial protection	个SOD and GSH-Px activity	
	Leuconostoc mesenteroides	↓Neuronal apoptosis	\downarrow NF-κB/MAPK signaling pathway	
	Weissella cibaria	↑Axonal regeneration	Dectin-1-mediated cytokine production	
	Candida, Saccharomyces,			
	Aspergillus			
Equol	Coriobacteriaceae	\downarrow Neuroinflammation	\downarrow NF- κ B/MAPK-mediated transcription of	47, 96, 97,
	Adlercreutzia equolifaciens	↓Neuronal apoptosis	inflammatory mediators	100
	Slackia equolifaciens	Cytoprotective and anti-inflammatory effects	\downarrow Microglia activation	
	Lactococcus garvieae	↑Neurite outgrowth	↑Neurotrophins production through	
	Lactobacillus intestinalis	Modulation of synaptic plasticity and cognitive	astrocytes	
	Bifidobacterium breve	functions	Agonist of ERβ	
Urolithins	Gordonibacter urolithinfaciens	\downarrow Neuroinflammation	\downarrow Proinflammatory cytokines	98, 99, 101-
Jrolithins	Gordonibacter pamelaeae	↓Neuronal apoptosis	↑Anti-inflammatory cytokines	104, 138
Jrolithins	Gordonibucter punicidede			
Jrolithins	Ellagibacter isourolithinifaciens	Antioxidant and anti-inflammatory effects	\downarrow Activation of microglia, DCs and T cells	
Jrolithins		Antioxidant and anti-inflammatory effects ↑Integrity of BBB and intestinal barrier	$\sqrt{\text{Activation of microglia, DCs and T cells}}$ $\sqrt{\text{ROS production}}$	
Urolithins	Ellagibacter isourolithinifaciens		•	
Jrolithins	Ellagibacter isourolithinifaciens Lactococcus garvieae	\uparrow Integrity of BBB and intestinal barrier	↓ROS production	

Note: The directions of the effects and mechanisms are indicated by symbols as follows: increase/upregulation (\uparrow) and decrease/downregulation (\downarrow). 3-HAA: 3-Hydroxyanthranilic acid; 5-HTP: 5-hydroxytryptophan; AhR: aryl hydrocarbon receptor; AMPK: 5'-adenosine monophosphate-activated protein kinase; A β : β -amyloid; BBB: blood-brain barrier; BCFAs: branched-chain fatty acids; BDNF: brain-derived neurotrophic factor; CLA: conjugated linoleic acid; CLNA: conjugated linolenic acid; CNS: central nervous system; DCs: dendritic cells; ENS: enteric nervous system; EPSs: exopolysaccharides; ERK1: extracellular-signal-regulated kinase 1; ER β : estrogen receptor β ; FFARs: free fatty acid receptors; FXR: farnesoid X receptor; GABA: γ -aminobutyric acid; GAP-43: growth-associated protein 43; GPR30: G protein-coupled receptor 30; GSH-Px: glutathione peroxidase; iNOS: inducible nitric oxide synthase; MAPK: mitogen-activated protein kinase; NF-kB: nuclear factor kappa B; NGF: nerve growth factor alpha; Nrf2: nuclear factor-erythroid-derived 2-like 2; PGC-1 α : peroxisome proliferator-activated receptor agamma coactivator 1-alpha; PI3K: phosphatidylinositol 3-kinase; PPAR α : peroxisome proliferator-activated receptor alpha; ROS: reactive oxygen species; SIRT: sirtuin; SKN-1: skinhead 1; SOD: superoxide dismutase; TJ: tight junction.

Notably, SCFAs have also been shown to control the maturation and function of microglia, which are crucial cells involved in neuroinflammatory responses and the maintenance of brain homeostasis. An in vivo study with germ-free mice revealed that SCFAs can modulate the maturity and morphology of microglia, including the number of segments, branching points, terminal points and cell volumes.⁵² Another study demonstrated that butyrate can reduce LPS-induced microglial activation and the depressive state.⁵³ Taken together, these findings indicate that SCFAs play crucial roles in microglial maturation and activation, possibly through epigenetic regulation that involves the inhibition of histone deacetylases. Interestingly, SCFAs, e.g., valeric acid, interfere with β -amyloid (A β) peptide 1–40 and 1–42 fibril formation in vitro and prevent α-synuclein monomers from pairing and aggregating into fibrils.¹⁸ Given that microglia play crucial roles in clearing aggregated proteins and maintaining brain homeostasis, the ability of SCFAs to modulate protein aggregation may be somewhat explained by the influence of SCFAs on microglial function. Notably, SCFAs exhibit positive effects on health at low to moderate physiological concentrations. At these concentrations, SCFAs can reduce inflammation and promote the production of beneficial neurotrophic factors. However, at high concentrations, SCFAs can become detrimental, leading to increased inflammation and oxidative stress.⁵⁴

Lactic acid, typically produced by lactic acid bacteria such as *Lactobacillus, Lactococcus,* and *Leuconostoc,* influences the production of GABA, a neurotransmitter that reduces neuronal excitability and protects against excitotoxicity, a process that leads to neuronal injury and death due to excessive stimulation by neurotransmitters. Additionally, lactate can be converted by various bacterial species into SCFAs, contributing to the overall metabolite pool. In the brain, owing to its ability to be metabolized into glutamate, lactate serves as an energy substrate for neurons, supports synaptic plasticity, and is essential for memory formation.^{55, 56}

Several recent studies have provided comprehensive evidence that the gut microbiota can produce conjugated fatty acid metabolites from a fat-enriched diet. Conjugated fatty acids, such as conjugated linoleic acid (CLA) and conjugated linolenic acid, are known to exhibit anti-inflammatory and antioxidant activities.^{57, 58} Furthermore, CLA and conjugated linolenic acid can impact the redistribution of ZO-1 and occludin, thus affecting the gut barrier and BBB functions. CLA was also shown to enter the brain and suppress neuroinflammation and the accumulation of AB. The neuroprotective effects of CLA are mediated through the upregulation of the anti-inflammatory cytokine IL-10 in astrocytes, a process induced by the activation of the nuclear transcription factor peroxisome proliferator-activated receptor gamma.⁵⁹ CLA may also help improve the breakdown of eicosanoids and oxidative stress products by increasing peroxisomal β -oxidation. This process enhances the body's ability to fight inflammation and oxidative stress, making CLA an effective anti-inflammatory and antioxidant agent. Moreover, CLA exerts a neuroprotective effect against glutamate excitotoxicity in the primary culture of rodent cortical neurons.⁶⁰ The antioxidant properties of CLA, including its ability to reduce reactive oxygen species (ROS) and increase the expression of antioxidant enzymes such as superoxide dismutase and glutathione peroxidase, help prevent age-associated neuronal damage in a mouse neurodegeneration model.⁶¹

Gut bacteria, such as Bacteroides fragilis, Ruminococcus lactaris, Faecalibacterium prausnitzii, and Lactiplantibacillus plantarum, have been reported to produce a variety of vitamins, including thiamin (vitamin B1), riboflavin (vitamin B2), niacin (vitamin B3), pantothenic acid (vitamin B5), pyridoxine (vitamin B6), biotin (vitamin B7), folate (vitamin B9), cobalamin (vitamin B12) and menaquinone (vitamin K2).⁶²⁻⁶⁴ As enzymatic cofactors, group B vitamins play crucial roles in numerous physiological processes, including the metabolism of glucose, fatty acids, and amino acids; the tryptophan-kynurenine pathway; and the synthesis of various neurotransmitters and neurohormones, such as serotonin, dopamine, adrenaline, acetylcholine, GABA, glutamate, and melatonin.^{33, 65} Since B vitamins are key players in metabolic pathways, it is not surprising that they modulate mitochondrial function, myelin formation, the response to oxidative stress and neuroinflammation.⁶⁶ For example, the administration of the thiamine-producing L. plantarum CRL 1905 strain was shown to inhibit neuronal death in a murine Parkinson's disease model by decreasing the expression of proinflammatory cytokines.⁶⁷ The gut microbiota also plays a major role in synthesizing vitamin K2.³² This vitamin is essential for the synthesis of sphingolipids, which are components of myelin and cell membranes.⁶⁸ Vitamin K2 was shown to inhibit NF-kB signaling and the production of proinflammatory cytokines, such as IL-1 β and IL-6.⁶⁹ The antioxidant properties of vitamin K2 further increase its neuroprotective potential. It neutralizes free radicals and reduces oxidative stress, which is common in the pathogenesis of many neurodegenerative diseases.⁴⁴ In Parkinson's disease, treatment with vitamin K2 reduces the formation of α -synuclein fibrils and downregulates inducible nitric oxide synthase, thereby decreasing neuroinflammation and apoptosis. In Alzheimer's disease, vitamin K2 promotes sphingolipid formation while decreasing the formation of A β plaques, H₂O₂ cytotoxicity, ROS and apoptosis.⁶⁸ These actions collectively contribute to neuroprotection and the mitigation of disease progression.

Tryptophan metabolites, such as indole derivatives and 3-HAA, play a vital role in regulating neuroinflammation and oxidative stress through the kynurenine pathway.¹⁹ Indole derivatives can modulate the activity of microglia and astrocytes, thus reducing the production of proinflammatory cytokines. Indole, indoxyl-3-sulfate, indole-3-propionic acid and indole-3aldehyde can activate AhR signaling in astrocytes, thus suppressing neuroinflammation.⁷⁰ Microbiota-derived indoles have also been shown to inhibit microglial activation and the NF-kB signaling pathway in a mouse model of Alzheimer's disease, which in turn prevents A β accumulation and tau hyperphosphorylation. The same study has shown that indoles reverse injury to the gut barrier by increasing the expression of TJ proteins and improving the immunity of the host.⁷¹ In addition to its anti-inflammatory activity, indole-3-propionic acid also has ROS-scavenging and BBB-protective properties.^{25, 72} This metabolite was shown to reverse endothelial dysfunction by upregulating the expression of TJ proteins in endothelial cells.⁷³ Furthermore, indole-3-propionic acid protects hippocampal cornu ammonis area 1 neurons from ischemic damage and is suggested to modulate mitochondrial biogenesis in hippocampal interneurons through a mechanism mediated by peroxisome proliferator-activated receptor gamma coactivator 1-alpha.⁷⁴ Additionally, microbiotaderived indole metabolites can reduce oxidative stress, inflammation, and neuronal apoptosis via the G proteincoupled receptor 30/5'-adenosine monophosphateactivated protein kinase/Sirtuin 1 pathway.75 Another tryptophan metabolite, 3-HAA, also contributes to antioxidant defense by scavenging free radicals, thus protecting neurons from oxidative damage. 3-HAA reportedly enhances resistance to oxidative stress by activating the nuclear factor-erythroid-derived 2-like 2 (Nrf2)/Skinhead 1 signaling pathway, which promotes increased lifespan and reduced aging.⁷⁶ It was also demonstrated that 3-HAA can inhibit LPSinduced inflammation through the suppression of the phosphatidylinositol 3-kinase/NF-κB signaling

pathways.^{28, 77} Additionally, kynurenic acid, another tryptophan metabolite, exerts neuroprotective effects by modulating glutamate receptors, thus preventing excitotoxicity, which can damage neurons and impede neurogenesis.^{30, 78}

Deconjugated bile acids produced by host-gut microbial cometabolism have been reported to modulate lipid metabolism and reduce oxidative stress, providing neuroprotective health benefits.⁷⁹⁻⁸² These bile acids also influence signaling pathways that regulate inflammation and cell survival, further contributing to brain health. Moreover, bile acids in the bloodstream affect BBB permeability by impacting TJ proteins such as occludin.⁷ Deconjugated bile acids exhibit neurosteroid-like properties by interacting with receptors for neurotransmitters, including GABA and N-methyl-D-aspartate receptors. Their neuroprotective action may also be associated with the inhibition of AB accumulation, mitochondrial damage, and apoptosis.⁷⁹ Through their receptors in the brain and ENS, bile acids exert neuroendocrine effects via nuclear farnesoid X receptor, which is crucial for normal brain function, as evidenced by studies showing altered neurotransmitter levels, impaired cognitive function, and motor coordination upon farnesoid X receptor deletion.^{79, 83}

EPSs produced by beneficial gut bacteria demonstrate neuroprotective properties by functioning as antioxidants and modulating immune responses. Several studies have highlighted the antioxidant activity of EPSs produced by lactic acid bacteria, which effectively scavenge superoxide anions and hydroxyl radicals.84-87 EPSs also increase the activity of vital antioxidant enzymes such as catalase, superoxide dismutase, and glutathione peroxidase. For example, EPS from Lacticaseibacillus rhamnosus has been shown to mitigate oxidative stress and reduce inflammatory brain injury induced by D-galactose.⁸⁶ Moreover, EPSs reduce Aβ-mediated neurotoxicity in human neuroblastoma SH-SY5Y cells by decreasing apoptotic activity and stabilizing the mitochondrial membrane potential.84,85

Like gut bacteria, gut fungi can also produce EPSs. These complex carbohydrate polymers play crucial roles in the gut environment, contributing to the formation of biofilms, modulating immune responses, and influencing the gut microbial community.³⁸ For example, the gut mycobiome can produce the polysaccharide β -1,3-glucan, which interacts with Dectin-1, a well-known pattern recognition receptor expressed by microglia and dendritic cells.⁸⁸ This interaction modulates cytokine expression, which can cross the intestinal barrier and

the BBB, thereby influencing brain function. Notably, β -1,3-glucan has been shown to promote CNS axon regeneration in a Dectin-1-dependent manner, likely through its effects on innate immunity.⁸⁸ Furthermore, β-glucan derived from Saccharomyces cerevisiae has been demonstrated to improve cognitive dysfunction via the gut-brain axis. Studies have shown that the administration of β-glucan leads to enhanced cognitive function and reduced pathological changes, including decreased AB plaque deposition, in Alzheimer's disease models.^{89, 90} Additionally, β-glucan supplementation has been associated with beneficial alterations in the composition of the gut microbiota and the production of metabolites such as SCFAs. These changes contribute to the overall reduction in inflammatory factors and microglial activation in the cerebral cortex and hippocampus, ultimately controlling neuroinflammation and supporting brain health.⁹⁰

Gut fungi are known to produce other types of bioactive metabolites, such as the phenolic alcohol tyrosol and the sesquiterpene alcohol farnesol.³⁸ Tyrosol and farnesol are both quorum-sensing molecules that play crucial roles in fungal communication and pathogenicity. In the gut environment, these compounds modulate fungal growth, biofilm formation, and the host immune response.⁹¹ Although the effects of these compounds produced by gut fungi on the nervous system have not yet been documented, it is important to note that plant-derived tyrosol and farnesol are known as neuroprotective, antioxidant, and anti-inflammatory agents that can promote remyelination and synaptogenesis.⁹²⁻⁹⁵ It is plausible that these compounds produced by gut fungi might exert comparable benefits on the nervous system. However, further research is needed to elucidate the specific roles of gut fungus-derived tyrosol and farnesol within the microbiota-gut-brain axis.

Some gut microbiota genera have demonstrated the ability to metabolize dietary polyphenols to produce microbiota-derived secondary metabolites, such as equol and urolithins.⁹⁶⁻⁹⁹ This modification of polyphenolic compounds significantly increases their bioavailability, maximizing their absorption and potential health-promoting effects. For example, equol, a metabolite produced from soy isoflavones by certain gut bacteria, has shown estrogen-like neuroprotective effects.^{47, 96,100} It can reduce inflammation and oxidative damage in the brain, promote neuronal health and prevent the neurodegenerative changes associated with aging and disease. The antioxidant properties of equol are largely attributed to its interaction with estrogen receptor β, which activates the extracellularsignal-regulated kinase (ERK)/NF-κB signaling pathway, thereby influencing transcription, cytokine production, and cell survival.⁹⁷ The gut microbiota also metabolizes ellagic acid to produce bioactive secondary metabolites called urolithins.^{98, 99, 101} Among these, urolithin A is notable for its potent anti-inflammatory and antioxidant properties. It induces mitochondria-selective autophagy, which helps to protect mitochondrial function during aging.⁹⁹ Urolithins have demonstrated promising neuroprotective effects by alleviating the damage induced by Aβ25-35, reducing lactate dehydrogenase leakage, decreasing ROS production, inhibiting neuroinflammation and neuronal apoptosis, and promoting neurite outgrowth.¹⁰²⁻¹⁰⁴

ROLE OF GUT MICROBIOTA METABOLITES IN THE REGENERATION OF NERVOUS TISSUE

The neuroprotective properties of gut microbiota metabolites suggest their potential role in promoting the regeneration of nervous tissue. These metabolites are believed to play crucial roles in essential processes vital for regeneration, such as neurogenesis, axonal repair, and synaptogenesis.^{19, 20, 43} Furthermore, recent research underscores the pivotal role of these metabolites in supporting the growth and repair of neural structures.^{2, 44, 45, 67} The ability of microbial metabolites to increase neurogenesis and modulate synaptic plasticity highlights their multifaceted impact on nervous system recovery.²⁰

A variety of microbial metabolites have been reported to promote the neurogenesis and proliferation of glial cells. These metabolites interact with neural or glial progenitor cells, influencing their differentiation and proliferation. For example, enteric SCFAs at physiological concentrations were shown to promote the proliferation and mitosis of human neural progenitor cells and influence the expression of neurogenesis-related and apoptosis-related genes.¹⁰⁵ SCFAs can also modulate oligodendrocyte progenitor differentiation and myelination.¹³ The effect of SCFAs on oligodendrocyte progenitors can be explained by the impact of SCFAs on microglia and astrocytes.¹⁰⁶ Another metabolite that can influence the proliferation of neural progenitor cells is CLA. CLA was shown to promote the neuronal differentiation of rat neural stem cells, increase the number of Tuj-1-positive cells, and induce the proliferation of neurospheres derived from rat neural progenitor cells.⁶⁰ Tryptophan metabolites produced by the gut microbiota, such as indole derivatives and kynurenine pathway metabolites, also

play crucial roles in neurogenesis and brain repair. For example, indole was found to enhance neurogenesis in the hippocampus of adult mice, as indicated by an increased number of doublecortin-expressing cells in the dentate gyrus. This stimulatory effect on adult neurogenesis is mediated through metabolic- and immune-linked AhR signaling.¹⁰⁷ Tryptophan-derived indole-3-lactic acid, produced by Bifidobacterium, was found to activate AhR and promote nerve growth factor (NGF)-induced neuronal differentiation via the ERK pathway.²⁶ The ERK signaling pathway is crucial for protein translational regulation, particularly in the context of long-term potentiation, which is essential for synaptic plasticity and memory formation. This, along with the observed increase in neurite outgrowth, indicates that indole-3-lactic acid may be a promising modulator of neuronal development and synaptic function.

Additionally, microbial metabolites can influence the production and release of neurotrophic factors, such as brain-derived neurotrophic factor (BDNF), NGF and glial cell-derived neurotrophic factor.¹⁰⁸ These factors are critical for the survival, growth, and differentiation of neurons and play a significant role in brain repair and regeneration processes.²⁰ SCFAs, particularly butyrate, increase the expression of BDNF, a crucial factor for neuroplasticity, neuronal survival, and synaptic function.^{17, 51} Lactate can also induce BDNF expression in the hippocampus through the upregulation of SIRT deacetylase.^{7, 109} Furthermore, microbiota-derived indole-3-propionic acid was shown to induce the production of neuroprotective BDNF and NGF in neuronal cells by inhibiting the production of proinflammatory factors from microglia, which resulted in improved cognitive function and mental stress in elderly individuals.45

Microbial metabolites also play a significant role in axonal repair. Axons can be damaged by injury or disease, leading to loss of function and connectivity in the nervous system. SCFAs promote axonal regeneration by increasing the expression of genes involved in cytoskeletal dynamics and axonal growth.¹⁰⁸ Propionic acid, for example, upregulates the expression of growth-associated protein 43, a critical protein for axonal growth and regeneration. The antioxidative and neuroregenerative effects of propionate on Schwann cells and dorsal root ganglia are mediated by free fatty acid receptor 3-induced histone acetylase expression.⁴³ Lactic acid has also been shown to support axonal repair.⁷ Lactate serves as an energy source for neurons and glial cells, providing the necessary ATP for the metabolic demands of axonal regeneration. It can also act as an astrocyte signaling molecule and regulate neuronal functions, including synaptic plasticity and axonal integrity.¹¹⁰ Additionally, lactate can modulate the expression of genes involved in axonal growth and repair, further supporting the regeneration process.¹¹¹ Group B vitamins, which can be synthesized by various gut microorganisms, may also support axonal recovery and enhance neuronal function. Both demyelination and axonal deterioration are common in vitamin B deficiency as well as in many neurological diseases, including multiple sclerosis and Alzheimer's disease. Group B vitamins have been shown to enhance motor nerve regeneration and the recovery of muscle function in a rat model of peripheral nerve injury by reducing both Schwann cell decline and deterioration of the myelin sheath.¹¹² A recent study revealed that the gut metabolite indole-3-propionate promotes nerve regeneration and repair via an immune-mediated mechanism.¹⁹ This metabolite, synthesized by *Clostridium sporogenes*, plays a crucial role in facilitating axonal regeneration. Administering indole-3-propionate after sciatic injury significantly enhances axonal regeneration, thereby accelerating the recovery of sensory function. In terms of mechanism, RNA sequencing analysis of sciatic dorsal root ganglia revealed that neutrophil chemotaxis plays a role in the regenerative effects induced by indole-3-propionic acid.19

Microbial metabolites also influence myelination. For example, SCFAs have been reported to mitigate demyelination and promote myelin regeneration in the corpus callosum.¹³ Specifically, butyrate has been demonstrated to have a pro-regenerative effect on myelin in organotypic cerebellar slice cultures following lysolecithin-induced demyelination. Treatment with butyrate increased the number of mature Olig2⁺ CC-1⁺ oligodendrocytes.¹¹³ The effect of butyrate on remyelination can be explained by its role in inhibiting histone deacetylases, as studies have shown that pharmacological inhibitors of histone deacetylases can similarly enhance remyelination.¹³ Vitamin K2, which can be synthesized by the gut microbiota, also impacts the myelination of neurons. It is essential for the formation of sphingolipids, which are vital for neuronal growth and the myelination process within the nervous system.^{68, 114}

Synaptogenesis is another critical aspect of nervous tissue regeneration influenced by gut microbiota metabolites.⁷ SCFAs, particularly butyrate, can increase synaptogenesis and prevent dendritic spine loss and

neurite impairment. Butyrate treatment increases the percentage of mushroom-like spines and decreases the percentage of thin spines and stubby spines in the frontal cortex of obese mice.¹¹⁵ Mushroom-like spines with large spine heads form strong synaptic connections and have the longest lifetime, making them crucial for long-term memory storage, synaptic plasticity, and cognitive processes. Another microbiotaderived metabolite, indole, has been shown to impact synaptic maturation. Mice treated with indole presented elevated levels of synaptic markers, such as postsynaptic density protein 95 and synaptophysin, indicating that indole enhances synaptogenesis.¹⁰⁷ An independent study of Alzheimer's disease confirmed that indoles can increase the expression of synaptic proteins such as postsynaptic density protein 95 and synapsin I, thereby improving synaptic plasticity and cognitive impairment.⁷¹ In a mouse model of autism spectrum disorder, treatment with indole-3-propionic acid was shown to restore impaired inhibitory synaptic transmission in the hippocampal dentate gyrus via ERK1 signaling, resulting in improved social behavior.¹¹⁶ It has also been reported that indole-3-propionic acid plays an important role in synaptic pruning by preventing excessive synapse elimination and synaptic impairment. This neuroprotective mechanism involves the inhibition of microglial activation mediated by AhR.¹¹⁷

The effects of the gut microbiota on neuroregenerative processes can also be mediated by the ability of the gut microbiota to synthesize neurotransmitters or neurotransmitter precursors, such as GABA, serotonin, 5-hydroxytryptophan, tryptamine, dopamine and norepinephrine. The gut microbiota has been reported to regulate the biosynthesis of host serotonin, also known as 5-HT. Indigenous spore-forming bacteria from mouse and human microbiota promote the production of 5-HT from colonic enterochromaffin cells.⁸ 5-HT profoundly affects enteric neurons. By enhancing 5-HT production, the gut microbiota can influence synaptic signaling within the ENS, promoting better coordination of muscle contractions and overall GIT motility.¹¹⁸ Furthermore, the modulation of serotonin levels by gut bacteria may contribute to the protection and regeneration of enteric neurons, supporting the resilience and adaptability of the ENS in response to physiological and pathological challenges.¹¹⁹ Several bacterial genera, such as Lactobacillus, Lactococcus, Bifidobacterium, Bacteroides and Akkermansia, have been shown to produce GABA through the glutamate decarboxylase pathway.^{9, 120} GABA-producing bacteria were shown to modulate sensory neuron activity in

a rat model of visceral hypersensitivity, suggesting their potential as future treatments for recurrent abdominal pain and functional bowel disorders.^{66, 121} The involvement of the GABAergic system in the pathogenesis of mood disorders is now widely acknowledged. The microbiota can alter the expression of GABA receptors in the brain, thereby reducing anxiety and depression.⁴⁶ Additionally, GABA plays a critical role in the ENS, regulates motility and mucosal function, and affects the gut-brain axis through the vagus nerve.¹²² Dopamine and norepinephrine are another set of neurotransmitters whose metabolism is influenced by the gut microbiota.^{66, 119, 123} Thev plav pivotal roles in various aspects of brain function, including stress responses and motor control. However, the detailed mechanisms by which microbiota-derived dopamine and norepinephrine affect the CNS and ENS have not yet been fully investigated.

Therapeutic potential of gut microbiota metabolites

The nervous system has a limited ability to repair itself, making the development of strategies for neural regeneration and neuroprotection among the top priorities in neuroscience research. Effective treatments could significantly mitigate the long-term effects of neurological disorders, improve patient outcomes and ease economic strain on healthcare systems. Given the frequent occurrence of neurological disorders and often unsatisfactory clinical outcomes, improving recovery remains a significant challenge. In this context, the emerging role of gut microbiota metabolites in neuroregeneration and neuroprotection represents a promising and innovative frontier.² Accumulating evidence suggests that metabolites produced by the gut microbiota play crucial roles in neural repair processes by supporting neurogenesis, synaptogenesis, and axonal growth.^{4, 19, 20, 24, 43, 45, 47, 67, 96, 107, 116, 117} For example, neurogenesis, which can be mediated by the gut microbiota through neuroactive compounds such as serotonin and GABA, is impaired in germ-free mice, which lack these microorganisms and consequently have fewer neurons in the ENS.² Moreover, microbial metabolites can promote regenerative processes through an immune-mediated mechanism. For example, microbiota-derived indole-3-propionic acid was shown to affect immune cells and modulate inflammatory responses, creating a supportive environment for the functional recovery of sensory axons.¹⁹

As research increasingly uncovers the profound effects of microbial metabolites on the nervous system, there is a surge of interest in exploring gut microbiota metabolite-derived therapeutics for neuroprotection and regeneration. This has led to a heightened focus on the therapeutic potential of probiotics, postbiotics, and metabiotics (Figure 3). Overall, health-promoting microbial metabolites can be applied in a variety of ways. They can be produced in the GIT by live microorganisms (probiotics), or they can be introduced directly as bioactive compounds resulting from microbial metabolic processes (postbiotics).¹²⁴ Another approach involves the use of metabiotics, which collectively refer to microbial metabolites, constituents of nonlive microorganisms, and signaling molecules with well-defined chemical compositions.¹²⁵ Probiotics are considered a traditional method for influencing the microbiota-gut-brain axis, and numerous studies have shown the potential of probiotics for alleviating neurodegenerative disorders, as well as promoting neurogenesis and synaptogenesis.46, 67, 125 The advantages of using probiotics include their safety, nonpathogenic nature, and ability to interact directly with the gut microbiota via quorum sensing. For probiotics to generate health-promoting compounds effectively, they must survive the harsh conditions of the GIT, such as low pH and high concentrations of bile salts, and demonstrate a strong ability to adhere to intestinal epithelial cells. Moreover, because each person has a distinct gut microbiota profile, dietary habits, and geographical influences, probiotics can produce different metabolites in varying amounts when used by different individuals. This variability can lead to heterogeneous effects on the microbiota-gutbrain axis among different people. Another limitation of probiotics is their potential user-unfriendliness for immunocompromised individuals, which can restrict

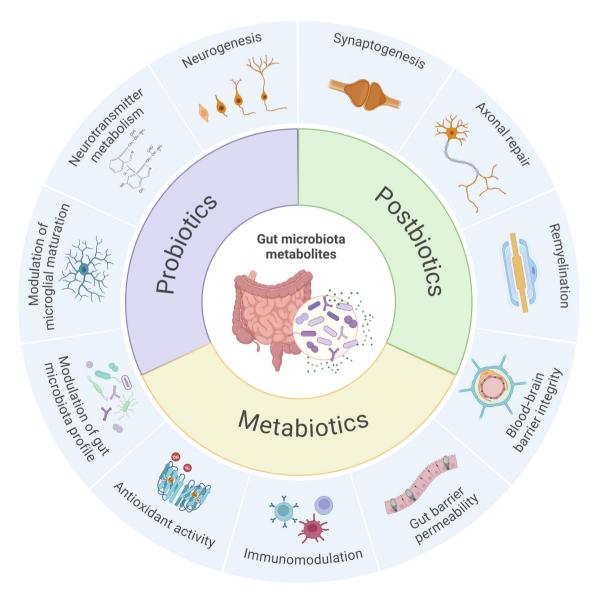


Figure 3: Schematic illustration of the therapeutic potential of gut microbiota metabolites Note: Created with BioRender.com.

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their use in therapies.¹²⁶ Alternative biotherapeutic products, such as postbiotics and metabiotics, which consist of microbial metabolites and microorganismderived cell components, may offer benefits over probiotics. They tend to be safer for sensitive populations, including neonates, young children, immunocompromised individuals, and critically ill patients, and are more resistant to the harsh conditions of the GIT.¹²⁷ Furthermore, controlling the production of metabolites-both in terms of quality and quantityis more straightforward under defined fermentation conditions with specific culture environments than in the host gut environment, ensuring more predictable, dosespecific physiological effects. Postbiotics and metabiotics also provide improved stability and transportability, which extends shelf-life and simplifies packaging.¹²⁵

Although many studies have shown the potential of microbial metabolites to promote neuroregenerative processes, the effectiveness and strategies of microbial metabolite therapies remain debatable, and more studies and large-scale clinical trials are needed. Moreover, it is essential to thoroughly investigate both the well-documented and potential species and possibly strains of human gut microbiota that could be used for the production of health-promoting metabolites. Future research should focus on identifying and characterizing the ability of these microorganisms to synthesize bioactive compounds, as well as understanding their functional roles within the microbiota-gut-brain axis. This study will help to uncover new strategies for developing targeted probiotic, postbiotic or metabiotic interventions, ultimately leading to advancements in maintaining nervous system health and broader therapeutic applications.

LIMITATIONS

This review has several limitations. Because this is a narrative review, it does not provide a systematic quantitative evaluation of the quality or relevance of all included studies, leading to potential selection bias and incomplete coverage of the literature. Moreover, the review focuses predominantly on bacterial metabolites, largely owing to the limited research available on fungal and archaeal metabolites. This emphasis may overshadow the potential impact of these less-studied metabolites on the microbiota–gut–brain axis, given the diverse nature of the gut microbiota. Given that this review addresses a wide array of microbial metabolites, the discussion of the mechanisms of action for each specific gut microbiota metabolite is limited in depth. Further research and clinical trials are needed before possibilities for evidence-based therapeutic applications of gut microbiota metabolites can be identified.

CONCLUSION AND FUTURE DIRECTIONS

Gut microbiota metabolites play crucial roles in maintaining the functional balance of the microbiotagut-brain axis, thus offering neuroprotection and promoting regeneration of nervous tissue. Research in this field continues, providing more information on specific mechanisms and approaches to repair neural tissue through microbiota modulation and the direct use of certain metabolites. Despite the significant therapeutic potential of these metabolites, further investigations are needed for evidence-based clinical applications. Key research directions include identifying probiotic strains that produce health-promoting metabolites and understanding the specific mechanisms through which the microbiota influences the nervous system. The challenge is elucidating the mechanisms by which microbial metabolites affect brain function, which stems from the unclear rate at which some microbial molecules are transported to the brain. It is also difficult to differentiate the direct effects of microbial metabolites on CNS function from those of indirect communication pathways, such as immunological pathways, or via the vagus nerve, which can confound in vivo studies. Therefore, further research is crucial to elucidate the mechanisms of absorption and transport of microbial metabolites and neurotransmitters from the gut to the brain. Additionally, exploring the roles of fungal and archaeal metabolites in the microbiota-gut-brain axis is important. Comprehensive experimental and clinical studies are needed to validate the protective effects of microbiota metabolites in neurological disorders.

Author contributions

Conceptualization, writing – original draft, and visualization: OK; conceptualization, writing – review & editing, and supervision: IL; conceptualization, writing – review & editing, and supervision: GS. All authors approved the final version of the manuscript.

Conflicts of interest

The authors declare no conflicts of interest.

Data availability statement

Not applicable.

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REFERENCES

- Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. *PLoS Biol*. 2016;14:e1002533.
- Herrera-Rincon C, Murciano-Brea J, Geuna S. Can we promote neural regeneration through microbiota-targeted strategies? Introducing the new concept of neurobiotics. *Neural Regen Res.* 2022;17:1965.
- 3. Calabrò S, Kankowski S, Cescon M, et al. Impact of gut microbiota on the peripheral nervous system in physiological, regenerative and pathological conditions. *Int J Mol Sci.* 2023;24:8061.
- 4. Loh JS, Mak WQ, Tan LKS, et al. Microbiota–gut–brain axis and its therapeutic applications in neurodegenerative diseases. *Sig Transduct Target Ther*. 2024;9:1-53.
- Kim CS. Roles of diet-associated gut microbial metabolites on brain health: cell-to-cell interactions between gut bacteria and the central nervous system. *Adv Nutr.* 2024;15:100136.
- Schroeder BO, Bäckhed F. Signals from the gut microbiota to distant organs in physiology and disease. *Nat Med.* 2016;22:1079-1089.
- Ahmed H, Leyrolle Q, Koistinen V, et al. Microbiota-derived metabolites as drivers of gut-brain communication. *Gut Microbes*. 2022;14:2102878.
- Yano JM, Yu K, Donaldson GP, et al. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell*. 2015;161:264-276.
- Strandwitz P, Kim KH, Terekhova D, et al. GABA-modulating bacteria of the human gut microbiota. *Nat Microbiol*. 2019;4:396-403.
- Dalile B, Van Oudenhove L, Vervliet B, Verbeke K. The role of short-chain fatty acids in microbiota–gut–brain communication. Nat Rev Gastroenterol Hepatol. 2019;16:461-478.
- Wang Y, Tong Q, Ma SR, et al. Oral berberine improves brain dopa/dopamine levels to ameliorate Parkinson's disease by regulating gut microbiota. *Signal Transduct Target Ther.* 2021;6:77.
- 12. Yang W, Cong Y. Gut microbiota-derived metabolites in the regulation of host immune responses and immune-related inflammatory diseases. *Cell Mol Immunol.* 2021;18:866-877.
- Sauma S, Casaccia P. Does the gut microbiota contribute to the oligodendrocyte progenitor niche? *Neurosci Lett*. 2020;715:134574.
- 14. Zhao YF, Wei DN, Tang Y. Gut microbiota regulate astrocytic functions in the brain: possible therapeutic consequences. *Curr Neuropharmacol.* 2021;19:1354-1366.
- 15. D'Alessandro G, Marrocco F, Limatola C. Microglial cells: Sensors for neuronal activity and microbiota-derived molecules. *Front Immunol.* 2022;13:1011129.
- Braniste V, Al-Asmakh M, Kowal C, et al. The gut microbiota influences blood-brain barrier permeability in mice. *Sci Transl Med.* 2014;6:263ra158.
- 17. Mansuy-Aubert V, Ravussin Y. Short chain fatty acids: the messengers from down below. *Front Neurosci*. 2023;17:1197759.
- Ahmed S, Busetti A, Fotiadou P, et al. In vitro characterization of gut microbiota-derived bacterial strains with neuroprotective properties. *Front Cell Neurosci.* 2019;13:402.
- 19. Serger E, Luengo-Gutierrez L, Chadwick JS, et al. The gut metabolite indole-3 propionate promotes nerve regeneration and repair. *Nature*. 2022;607:585-592.

- Glinert A, Turjeman S, Elliott E, Koren O. Microbes, metabolites and (synaptic) malleability, oh my! The effect of the microbiome on synaptic plasticity. *Biol Rev Camb Philos Soc.* 2022;97:582-599.
- 21. Han Y, Wang B, Gao H, et al. Vagus nerve and underlying impact on the gut microbiota-brain axis in behavior and neurodegenerative diseases. *J Inflamm Res.* 2022;15:6213-6230.
- 22. Geng ZH, Zhu Y, Li QL, Zhao C, Zhou PH. Enteric nervous system: the bridge between the gut microbiota and neurological disorders. *Front Aging Neurosci*. 2022;14:810483.
- 23. Silva YP, Bernardi A, Frozza RL. The role of short-chain fatty acids from gut microbiota in gut-brain communication. *Front Endocrinol (Lausanne)*. 2020;11:25.
- 24. Li H, Sun J, Wang F, et al. Sodium butyrate exerts neuroprotective effects by restoring the blood–brain barrier in traumatic brain injury mice. *Brain Res.* 2016;1642:70-78.
- 25. Konopelski P, Mogilnicka I. Biological effects of indole-3-propionic acid, a gut microbiota-derived metabolite, and its precursor tryptophan in mammals' health and disease. *Int J Mol Sci.* 2022;23:1222.
- Wong CB, Tanaka A, Kuhara T, Xiao JZ. Potential effects of indole-3-lactic acid, a metabolite of human bifidobacteria, on NGFinduced neurite outgrowth in PC12 cells. *Microorganisms*. 2020;8:398.
- 27. Chen Y, Xu J, Chen Y. Regulation of neurotransmitters by the gut microbiota and effects on cognition in neurological disorders. *Nutrients*. 2021;13:2099.
- Wang J, Zhu N, Su X, Gao Y, Yang R. Gut-microbiota-derived metabolites maintain gut and systemic immune homeostasis. *Cells*. 2023;12:793.
- 29. Parker A, Fonseca S, Carding SR. Gut microbes and metabolites as modulators of blood–brain barrier integrity and brain health. *Gut Microbes*. 2019;11:135-157.
- Shaw C, Hess M, Weimer BC. Microbial-derived tryptophan metabolites and their role in neurological disease: anthranilic acid and anthranilic acid derivatives. *Microorganisms*. 2023;11:1825.
- 31. Uebanso T, Shimohata T, Mawatari K, Takahashi A. Functional roles of B-vitamins in the gut and gut microbiome. *Mol Nutr Food Res.* 2020;64:e2000426.
- 32. Bonaldo F, Leroy F. Review: Bacterially produced vitamin K2 and its potential to generate health benefits in humans. *Trends Food Sci Technol.* 2024;147:104461.
- Rudzki L, Stone TW, Maes M, Misiak B, Samochowiec J, Szulc A. Gut microbiota-derived vitamins - underrated powers of a multipotent ally in psychiatric health and disease. *Prog Neuropsychopharmacol Biol Psychiatry*. 2021;107:110240.
- 34. Hou K, Wu ZX, Chen XY, et al. Microbiota in health and diseases. *Signal Transduct Target Ther.* 2022;7:135.
- Parte AC, Sardà Carbasse J, Meier-Kolthoff JP, Reimer LC, Göker M. List of Prokaryotic names with Standing in Nomenclature (LPSN) moves to the DSMZ. *Int J Syst Evol Microbiol*. 2020;70:5607-5612.
- Swer NM, Venkidesh BS, Murali TS, Mumbrekar KD. Gut microbiota-derived metabolites and their importance in neurological disorders. *Mol Biol Rep.* 2023;50:1663-1675.
- Enaud R, Vandenborght LE, Coron N, et al. The mycobiome: a neglected component in the microbiota-gut-brain axis. *Microorganisms*. 2018;6:22.

- Chin VK, Yong VC, Chong PP, Amin Nordin S, Basir R, Abdullah M. Mycobiome in the gut: a multiperspective review. *Mediators Inflamm.* 2020;2020:9560684.
- 39. Belvoncikova P, Splichalova P, Videnska P, Gardlik R. The human mycobiome: colonization, composition and the role in health and disease. *J Fungi (Basel)*. 2022;8:1046.
- 40. Kim JY, Whon TW, Lim MY, et al. The human gut archaeome: identification of diverse haloarchaea in Korean subjects. *Microbiome*. 2020;8:114.
- 41. Cai M, Kandalai S, Tang X, Zheng Q. Contributions of humanassociated archaeal metabolites to tumor microenvironment and carcinogenesis. *Microbiol Spectr.* 2022;10:e0236721.
- 42. Carrera I, Cacabelos R. Current drugs and potential future neuroprotective compounds for Parkinson's disease. *Curr Neuropharmacol.* 2019;17:295-306.
- 43. Grüter T, Mohamad N, Rilke N, et al. Propionate exerts neuroprotective and neuroregenerative effects in the peripheral nervous system. *Proc Natl Acad Sci U S A*. 2023;120:e2216941120.
- 44. Chatterjee K, Mazumder PM, Sarkar SR, et al. Neuroprotective effect of Vitamin K2 against gut dysbiosis associated cognitive decline. *Physiol Behav.* 2023;269:114252.
- Kim CS, Jung S, Hwang GS, Shin DM. Gut microbiota indole-3-propionic acid mediates neuroprotective effect of probiotic consumption in healthy elderly: A randomized, double-blind, placebo-controlled, multicenter trial and in vitro study. *Clin Nutr.* 2023;42:1025-1033.
- 46. Jia L, Xiao L, Fu Y, et al. Neuroprotective effects of probiotics on anxiety- and depression-like disorders in stressed mice by modulating tryptophan metabolism and the gut microbiota. *Food Funct*. 2024;15:2895-2905.
- Johnson SL, Park HY, Vattem DA, Grammas P, Ma H, Seeram NP. Equol, a blood–brain barrier permeable gut microbial metabolite of dietary isoflavone daidzein, exhibits neuroprotective effects against neurotoxins induced toxicity in human neuroblastoma SH-SY5Y Cells and Caenorhabditis elegans. *Plant Foods Hum Nutr.* 2020;75:512-517.
- 48. Fusco W, Lorenzo MB, Cintoni M, et al. Short-chain fattyacid-producing bacteria: key components of the human gut microbiota. *Nutrients*. 2023;15:2211.
- 49. Kopczyńska J, Kowalczyk M. The potential of short-chain fatty acid epigenetic regulation in chronic low-grade inflammation and obesity. *Front Immunol.* 2024;15:1380476.
- 50. Rose S, Bennuri SC, Davis JE, et al. Butyrate enhances mitochondrial function during oxidative stress in cell lines from boys with autism. *Transl Psychiatry*. 2018;8:42.
- 51. Mirzaei R, Bouzari B, Hosseini-Fard SR, et al. Role of microbiotaderived short-chain fatty acids in nervous system disorders. *Biomed Pharmacother*. 2021;139:111661.
- 52. Erny D, de Angelis ALH, Jaitin D, et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat Neurosci.* 2015;18:965-977.
- 53. Yamawaki Y, Yoshioka N, Nozaki K, et al. Sodium butyrate abolishes lipopolysaccharide-induced depression-like behaviors and hippocampal microglial activation in mice. *Brain Res.* 2018;1680:13-38.
- 54. O'Riordan KJ, Collins MK, Moloney GM, et al. Short chain fatty acids: Microbial metabolites for gut-brain axis signalling. *Mol Cell Endocrinol.* 2022;546:111572.

- 55. Chen X, Zhang Y, Wang H, Liu L, Li W, Xie P. The regulatory effects of lactic acid on neuropsychiatric disorders. *Discov Ment Health*. 2022;2:8.
- 56. Caspani G, Kennedy S, Foster JA, Swann J. Gut microbial metabolites in depression: understanding the biochemical mechanisms. *Microb Cell.* 2019;6:454-481.
- 57. Mei Y, Chen H, Yang B, Zhao J, Zhang H, Chen W. Linoleic acid triggered a metabolomic stress condition in three species of bifidobacteria characterized by different conjugated linoleic acidproducing abilities. J Agric Food Chem. 2021;69:11311-11321.
- Nasrollahzadeh A, Mollaei Tavani S, Arjeh E, Jafari SM. Production of conjugated linoleic acid by lactic acid bacteria; important factors and optimum conditions. *Food Chem X*. 2023;20:100942.
- 59. Fujita Y, Kano K, Kishino S, et al. Dietary cis-9, trans-11conjugated linoleic acid reduces amyloid β-protein accumulation and upregulates anti-inflammatory cytokines in an Alzheimer's disease mouse model. *Sci Rep.* 2021;11:9749.
- Murru E, Carta G, Manca C, et al. Conjugated linoleic acid and brain metabolism: a possible anti-neuroinflammatory role mediated by PPARα activation. *Front Pharmacol.* 2021;11:587140.
- 61. Monaco A, Ferrandino I, Boscaino F, et al. Conjugated linoleic acid prevents age-dependent neurodegeneration in a mouse model of neuropsychiatric lupus via the activation of an adaptive response. *J Lipid Res.* 2018;59:48-57.
- 62. Yoshii K, Hosomi K, Sawane K, Kunisawa J. Metabolism of dietary and microbial vitamin b family in the regulation of host immunity. *Front Nutr.* 2019;6:48.
- 63. Hossain KS, Amarasena S, Mayengbam S. B vitamins and their roles in gut health. *Microorganisms*. 2022;10:1168.
- 64. Zhan Q, Wang R, Thakur K, et al. Unveiling of dietary and gutmicrobiota derived B vitamins: Metabolism patterns and their synergistic functions in gut-brain homeostasis. *Crit Rev Food Sci Nutr. 2024;64:4046-4058.*
- 65. Roth W, Mohamadzadeh M. Vitamin B12 and gut-brain homeostasis in the pathophysiology of ischemic stroke. *EBioMedicine*. 2021;73:103676.
- 66. Miri S, Yeo J, Abubaker S, Hammami R. Neuromicrobiology, an emerging neurometabolic facet of the gut microbiome? *Front Microbiol.* 2023;14:1098412.
- Teran MDM, Perez Visñuk D, Savoy de Giori G, de Moreno de LeBlanc A, LeBlanc JG. Neuroprotective effect of thiamineproducing lactic acid bacteria in a murine Parkinsonian model. *Food Funct*. 2022;13:8056-8067.
- 68. Chatterjee K, Mazumder PM, Banerjee S. Vitamin K: a potential neuroprotective agent. *Rev Bras Farmacogn*. 2023;33:676-687.
- 69. Popescu A, German M. Vitamin K2 holds promise for Alzheimer's prevention and treatment. *Nutrients*. 2021;13:2206.
- Rothhammer V, Mascanfroni ID, Bunse L, et al. Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nat Med*. 2016;22:586-597.
- Sun J, Zhang Y, Kong Y, et al. Microbiota-derived metabolite Indoles induced aryl hydrocarbon receptor activation and inhibited neuroinflammation in APP/PS1 mice. *Brain Behav Immun*. 2022;106:76-88.
- Negatu DA, Gengenbacher M, Dartois V, Dick T. Indole propionic acid, an unusual antibiotic produced by the gut microbiota, with anti-inflammatory and antioxidant properties. *Front Microbiol.* 2020;11:575586.

- Wang Q, Lv H, Ainiwan M, et al. Untargeted metabolomics identifies indole-3-propionic acid to relieve Ang II-induced endothelial dysfunction in aortic dissection. *Mol Cell Biochem*. 2024;479:1767-1786.
- 74. Zhou X, Wu X, Wu Y, et al. Indole-3-propionic acid, a gut microbiota metabolite, protects against the development of postoperative delirium. *Ann Surg.* 2023;278:e1164-1174.
- Yin J, Zhang Y, Liu X, et al. Gut microbiota-derived indole derivatives alleviate neurodegeneration in aging through activating GPR30/AMPK/SIRT1 pathway. *Mol Nutr Food Res.* 2023;67:e2200739.
- 76. Dang H, Castro-Portuguez R, Espejo L, et al. On the benefits of the tryptophan metabolite 3-hydroxyanthranilic acid in Caenorhabditis elegans and mouse aging. *Nat Commun.* 2023;14:8338.
- Lee K, Kwak JH, Pyo S. Inhibition of LPS-induced inflammatory mediators by 3-hydroxyanthranilic acid in macrophages through suppression of PI3K/NF-κB signaling pathways. *Food Funct*. 2016;7:3073-3082.
- Sathyasaikumar KV, Blanco-Ayala T, Zheng Y, et al. The tryptophan metabolite indole-3-propionic acid raises kynurenic acid levels in the rat brain in vivo. *Int J Tryptophan Res*. 2024;17:11786469241262876.
- 79. Baloni P, Funk CC, Yan J, et al. Metabolic network analysis reveals altered bile acid synthesis and metabolism in Alzheimer's disease. *Cell Rep Med*. 2020;1:100138.
- Ayyash MM, Abdalla AK, AlKalbani NS, et al. Invited review: Characterization of new probiotics from dairy and nondairy products—Insights into acid tolerance, bile metabolism and tolerance, and adhesion capability. *J Dairy Sci.* 2021;104:8363-8379.
- 81. Mulak A. Bile acids as key modulators of the brain-gut-microbiota axis in Alzheimer's disease. *J Alzheimers Dis*. 2021;84:461-477.
- 82. Kiriyama Y, Nochi H. The biosynthesis, signaling, and neurological functions of bile acids. *Biomolecules*. 2019;9:232.
- Monteiro-Cardoso VF, Corlianò M, Singaraja RR. Bile acids: a communication channel in the gut-brain axis. *Neuromol Med*. 2021;23:99-117.
- Sirin S, Aslim B. Characterization of lactic acid bacteria derived exopolysaccharides for use as a defined neuroprotective agent against amyloid beta1–42-induced apoptosis in SH-SY5Y cells. *Sci Rep.* 2020;10:8124. doi:10.1038/s41598-020-65147-1
- Sirin S, Aslim B. Protective effect of exopolysaccharides from lactic acid bacteria against amyloid beta1-42induced oxidative stress in SH-SY5Y cells: Involvement of the AKT, MAPK, and NF-κB signaling pathway. *Process Biochem*. 2021;106:50-59.
- Kumari M, Dasriya VL, Nataraj BH, Nagpal R, Behare PV. Lacticaseibacillus rhamnosus-derived exopolysaccharide attenuates d-galactose-induced oxidative stress and inflammatory brain injury and modulates gut microbiota in a mouse model. *Microorganisms*. 2022;10:2046.
- Sørensen HM, Rochfort KD, Maye S, et al. Exopolysaccharides of lactic acid bacteria: production, purification and health benefits towards functional food. *Nutrients*. 2022;14:2938.
- Baldwin KT, Carbajal KS, Segal BM, Giger RJ. Neuroinflammation triggered by β-glucan/dectin-1 signaling enables CNS axon regeneration. *Proc Natl Acad Sci U S A*. 2015;112:2581-2586.
- Xu M, Mo X, Huang H, et al. Yeast β-glucan alleviates cognitive deficit by regulating gut microbiota and metabolites in Aβ1–42induced AD-like mice. *Int J Biol Macromol.* 2020;161:258-270.

- Zhang Q, Zhao W, Hou Y, et al. β-Glucan attenuates cognitive impairment of APP/PS1 mice via regulating intestinal flora and its metabolites. CNS Neurosci Ther. 2023;29:1690-1704.
- 91. Rodrigues CF, Černáková L. Farnesol and tyrosol: secondary metabolites with a crucial quorum-sensing role in candida biofilm development. *Genes*. 2020;11:444.
- 92. Taniguchi K, Yamamoto F, Arai T, et al. Tyrosol reduces amyloid-β oligomer neurotoxicity and alleviates synaptic, oxidative, and cognitive disturbances in Alzheimer's disease model mice. J Alzheimers Dis. 2019;70:937-952.
- De La Cruz JP, Ruiz-Moreno MI, Guerrero A, et al. Differences in the neuroprotective effect of orally administered virgin olive oil (olea europaea) polyphenols tyrosol and hydroxytyrosol in rats. J Agric Food Chem. 2015;63:5957-5963.
- 94. Doyle WJ, Walters D, Shi X, et al. Farnesol brain transcriptomics in CNS inflammatory demyelination. *Clin Immunol.* 2023;255:109752.
- 95. Sell LB, Ramelow CC, Kohl HM, et al. Farnesol induces protection against murine CNS inflammatory demyelination and modifies gut microbiome. *Clin Immunol.* 2022;235:108766.
- 96. Subedi L, Ji E, Shin D, Jin J, Yeo JH, Kim SY. Equol, a dietary daidzein gut metabolite attenuates microglial activation and potentiates neuroprotection in vitro. *Nutrients*. 2017;9:207.
- 97. Mayo B, Vázquez L, Flórez AB. Equol: a bacterial metabolite from the daidzein isoflavone and its presumed beneficial health effects. *Nutrients*. 2019;11:2231.
- 98. García-Villalba R, Beltrán D, Frutos MD, Selma MV, Espín JC, Tomás-Barberán FA. Metabolism of different dietary phenolic compounds by the urolithin-producing human-gut bacteria Gordonibacter urolithinfaciens and Ellagibacter isourolithinifaciens. *Food Funct*. 2020;11:7012-7022.
- 99. Mi H, Liu S, Hai Y, et al. Lactococcus garvieae FUA009, a novel intestinal bacterium capable of producing the bioactive metabolite urolithin a from ellagic acid. *Foods*. 2022;11:2621.
- 100. Moran LM, McLaurin KA, Booze RM, Mactutus CF. Neurorestoration of sustained attention in a model of HIV-1 associated neurocognitive disorders. *Front Behav Neurosci.* 2019;13:169.
- 101. He F, Bian Y, Zhao Y, et al. In vitro conversion of ellagic acid to urolithin A by different gut microbiota of urolithin metabotype A. *Appl Microbiol Biotechnol.* 2024;108:215.
- 102. Chen P, Chen F, Lei J, Li Q, Zhou B. Activation of the miR-34a-mediated SIRT1/mTOR signaling pathway by urolithin a attenuates d-galactose-induced brain aging in mice. *Neurotherapeutics*. 2019;16:1269-1282.
- 103. Lin XH, Ye XJ, Li QF, et al. Urolithin A prevents focal cerebral ischemic injury via attenuating apoptosis and neuroinflammation in mice. *Neuroscience*. 2020;448:94-106.
- 104. Toney AM, Albusharif M, Works D, et al. Differential Effects of Whole Red Raspberry Polyphenols and Their Gut Metabolite Urolithin A on Neuroinflammation in BV-2 Microglia. *Int J Environ Res Public Health*. 2020;18:68.
- 105. Yang LL, Millischer V, Rodin S, MacFabe DF, Villaescusa JC, Lavebratt C. Enteric short-chain fatty acids promote proliferation of human neural progenitor cells. *J Neurochem*. 2020;154:635-646.
- 106. Gao Y, Xie D, Wang Y, Niu L, Jiang H. Short-chain fatty acids reduce oligodendrocyte precursor cells loss by inhibiting the activation of astrocytes via the SGK1/IL-6 signalling pathway. *Neurochem Res.* 2022;47:3476-3489.

- 107. Wei GZ, Martin KA, Xing PY, et al. Tryptophan-metabolizing gut microbes regulate adult neurogenesis via the aryl hydrocarbon receptor. *Proc Natl Acad Sci U S A.* 2021;118:e2021091118.
- 108. Lan Z, Tang X, Lu M, Hu Z, Tang Z. The role of short-chain fatty acids in central nervous system diseases: A bibliometric and visualized analysis with future directions. *Heliyon*. 2024;10:e26377.
- 109. El Hayek L, Khalifeh M, Zibara V, et al. Lactate mediates the effects of exercise on learning and memory through SIRT1dependent activation of hippocampal brain-derived neurotrophic factor (BDNF). *J Neurosci.* 2019;39:2369-2382.
- 110. Hemati-Gourabi M, Cao T, Romprey MK, Chen M. Capacity of astrocytes to promote axon growth in the injured mammalian central nervous system. *Front Neurosci.* 2022;16:955598.
- 111. Li F, Sami A, Noristani HN, et al. Glial metabolic rewiring promotes axon regeneration and functional recovery in the central nervous system. *Cell Metab*. 2020;32(5):767-785.e7.
- 112. Mandić M, Mitić K, Nedeljković P, et al. Vitamin B complex and experimental autoimmune encephalomyelitis –Attenuation of the clinical signs and gut microbiota dysbiosis. *Nutrients*. 2022;14(6):1273.
- 113. Chen T, Noto D, Hoshino Y, Mizuno M, Miyake S. Butyrate suppresses demyelination and enhances remyelination. J *Neuroinflammation*. 2019;16:165.
- 114. Zhang T, O'Connor C, Sheridan H, Barlow JW. Vitamin K2 in Health and Disease: A Clinical Perspective. *Foods*. 2024;13(11):1646. doi:10.3390/foods13111646
- 115. Ge X, Zheng M, Hu M, et al. Butyrate ameliorates quinolinic acid–induced cognitive decline in obesity models. *J Clin Invest.* 2023;133:e154612.
- 116. Jiang J, Wang D, Jiang Y, et al. The gut metabolite indole-3propionic acid activates ERK1 to restore social function and hippocampal inhibitory synaptic transmission in a 16p11.2 microdeletion mouse model. *Microbiome*. 2024;12:66.
- 117. Wang T, Chen B, Luo M, et al. Microbiota-indole 3-propionic acidbrain axis mediates abnormal synaptic pruning of hippocampal microglia and susceptibility to ASD in IUGR offspring. *Microbiome*. 2023;11:245.
- 118. Bellono NW, Bayrer JR, Leitch DB, et al. Enterochromaffin cells are gut chemosensors that couple to sensory neural pathways. *Cell*. 2017;170(1):185-198.e16. doi:10.1016/j.cell.2017.05.034
- 119. Dicks LMT. Gut bacteria and neurotransmitters. *Microorganisms*. 2022;10:1838.
- 120. Konstanti P, Ligthart K, Fryganas C, et al. Physiology of γ-aminobutyric acid production by Akkermansia muciniphila. *Appl Environ Microbiol*. 2024;90:e0112123.
- 121. Laroute V, Beaufrand C, Gomes P, et al. Lactococcus lactis NCDO2118 exerts visceral antinociceptive properties in rat via GABA production in the gastro-intestinal tract. *Elife*. 2022;11:e77100.
- 122. Braga JD, Thongngam M, Kumrungsee T. Gamma-aminobutyric acid as a potential postbiotic mediator in the gut–brain axis. *NPJ Sci Food*. 2024;8:16.
- 123. Maini Rekdal V, Bess EN, Bisanz JE, Turnbaugh PJ, Balskus EP. Discovery and inhibition of an interspecies gut bacterial pathway for Levodopa metabolism. *Science*. 2019;364:eaau6323.

- 124. Żółkiewicz J, Marzec A, Ruszczyński M, Feleszko W. Postbiotics— A step beyond pre- and probiotics. *Nutrients*. 2020;12:2189.
- 125. Jang HJ, Lee NK, Paik HD. A narrative review on the advance of probiotics to metabiotics. *J Microbiol Biotechnol*. 2024;34:487-494.
- 126. Ma L, Tu H, Chen T. Postbiotics in human health: a narrative review. *Nutrients*. 2023;15:291.
- 127. Hijová E. Postbiotics as metabolites and their biotherapeutic potential. *Int J Mol Sci.* 2024;25:5441.
- 128. Rios-Covian D, González S, Nogacka AM, et al. An overview on fecal branched short-chain fatty acids along human life and as related with body mass index: associated dietary and anthropometric factors. *Front Microbiol*. 2020;11:973.
- 129. O'Hagan C, Li JV, Marchesi JR, Plummer S, Garaiova I, Good MA. Long-term multi-species *Lactobacillus* and *Bifidobacterium* dietary supplement enhances memory and changes regional brain metabolites in middle-aged rats. *Neurobiol Learn Mem.* 2017;144:36-47.
- 130. Abd Mutalib N, Syed Mohamad SA, Jusril NA, Hasbullah NI, Mohd Amin MCI, Ismail NH. Lactic acid bacteria (LAB) and neuroprotection, what is new? An up-to-date systematic review. *Pharmaceuticals (Basel)*. 2023;16:712.
- 131. Ma T, Shen X, Shi X, et al. Targeting gut microbiota and metabolism as the major probiotic mechanism An evidence-based review. *Trends Food Sci Technol.* 2023;138:178-198.
- 132. Steiner J, Dobrowolny H, Guest PC, et al. Plasma anthranilic acid and leptin levels predict HAM-D scores in depressed women. *Int J Tryptophan Res.* 2021;14:11786469211016474.
- 133. Laroute V, Aubry N, Audonnet M, Mercier-Bonin M, Daveran-Mingot ML, Cocaign-Bousquet M. Natural diversity of lactococci in γ-aminobutyric acid (GABA) production and genetic and phenotypic determinants. *Microb Cell Fact.* 2023;22:178.
- 134. Chen M, Ruan G, Chen L, et al. Neurotransmitter and intestinal interactions: focus on the microbiota-gut-brain axis in irritable bowel syndrome. *Front Endocrinol (Lausanne).* 2022;13:817100.
- 135. Hamamah S, Aghazarian A, Nazaryan A, Hajnal A, Covasa M. Role of microbiota-gut-brain axis in regulating dopaminergic signaling. *Biomedicines*. 2022;10:436.
- 136. Ojeda J, Ávila A, Vidal PM. Gut microbiota interaction with the central nervous system throughout life. J Clin Med. 2021;10:1299.
- 137. Jayachandran M, Chen J, Chung SSM, Xu B. A critical review on the impacts of β -glucans on gut microbiota and human health. *J Nutr Biochem*. 2018;61:101-110.
- 138. Li HL, Zhang SY, Ren YS, et al. Identification of ellagic acid and urolithins as natural inhibitors of Aβ25–35-induced neurotoxicity and the mechanism predication using network pharmacology analysis and molecular docking. *Front Nutr.* 2022;9:966276.

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