SUPPLEMENTARY MATERIAL

Table S1. Mutation rates calculated for specific mutation types in the URA3 sequence in the rev3Δ msh6Δ background. Mutation spectra for strains OR1,

OR2, pol3-L612M OR1, pol3-L612M OR2, pol2-M644G OR1, and pol2-M644G OR2 were presented previously in [44] and [50].

	OF	R1	OF	R2	dpb2 Ol	2 <i>-100</i> R1	dpb2 Ol	2-100 R2	pol3-l O	L612M R1	pol3-l O	L612M R2	dpb2 pol3-l Ol	2-100 L612M R1	dpt pol3 C	02-100 -L612M 0R2	pol2-l Ol	<b>//644G</b> R1	pol2-l O	//644G R2	dpb2 pol2-l O	2-100 M644G R1	dpb2 pol2-l O	2-100 M644G R2
Transitions	103ª	1.10 <sup>⊳</sup>	37	0.53	80	9.49	67	7.54	161	35.42	112	22.76	119	59.73	96	76.03	83	28.09	68	26.12	98	52.30	84	62.61
T→C	23	0.25	5	0.07	11	1.31	18	2.03	53	11.66	2	0.41	27	13.55	6	4.75	6	2.03	17	6.53	13	6.94	18	13.42
<i>T</i> →C at 97 °	7	0.07	1	0.01	1	0.12	2	0.23	24	5.28	0	0.00	8	4.02	2	1.58	2	0.68	5	1.92	5	2.67	5	3.73
<i>T</i> →C at OS <sup>d</sup>	16	0.17	4	0.06	10	1.19	16	1.80	29	6.38	2	0.41	19	9.54	4	3.17	4	1.35	12	4.61	8	4.27	13	9.69
A→G	2	0.02	6	0.09	2	0.24	4	0.45	0	0.00	18	3.66	0	0.00	13	10.30	2	0.68	3	1.15	3	1.60	6	4.47
C→T	13	0.14	18	0.26	16	1.90	27	3.04	1	0.22	85	17.27	17	8.53	55	43.56	20	6.77	17	6.53	19	10.14	25	18.63
C→T at 310	4	0.04	11	0.16	5	0.59	11	1.24	1	0.22	43	8.74	5	2.51	40	31.68	6	2.03	9	3.46	4	2.13	15	11.18
C→T at OS	9	0.10	7	0.10	11	1.31	16	1.80	0	0.00	42	8.54	12	6.02	15	11.88	14	4.74	8	3.07	15	8.00	10	7.45
G→A	65	0.70	8	0.11	51	6.05	18	2.03	107	23.54	7	1.42	75	37.64	22	17.42	55	18.61	31	11.91	63	33.62	35	26.09
<i>G</i> →A at 764	18	0.19	2	0.03	8	0.95	2	0.23	45	9.90	2	0.41	15	7.53	3	2.38	10	3.38	9	3.46	12	6.40	4	2.98
$G \rightarrow A at OS$	47	0.50	6	0.09	43	5.10	16	1.80	62	13.64	5	1.02	60	30.11	19	15.05	45	15.23	22	8.45	51	27.22	31	23.11
Transversions	54	0.58	87	1.24	37	4.39	55	6.19	8	1.76	54	10.97	17	8.53	33	26.14	69	23.35	35	13.44	57	30.42	49	36.52
G→T	25	0.27	70	0.99	20	2.37	50	5.63	2	0.44	47	9.55	8	4.02	22	17.42	37	12.52	23	8.83	26	13.87	24	17.89
G→T at 679/706	8	0.09	44	0.63	6	0.71	13	1.46	0	0.00	9	1.83	3	1.51	5	3.96	8	2.71	16	6.15	15	8.00	11	8.20
$G \rightarrow T$ at OS	17	0.18	26	0.37	14	1.66	37	4.16	2	0.44	38	7.72	5	2.51	17	13.46	29	9.81	7	2.69	11	5.87	13	9.69
C→A	14	0.15	9	0.13	8	0.95	0	0.00	4	0.88	0	0.00	4	2.01	2	1.58	2	0.68	2	0.77	11	5.87	11	8.20
T→G	9	0.10	4	0.06	2	0.24	2	0.23	1	0.22	0	0.00	2	1.00	0	0.00	2	0.68	3	1.15	1	0.53	4	2.98
A→C	2	0.02	1	0.01	1	0.12	1	0.11	0	0.00	3	0.61	0	0.00	1	0.79	0	0.00	0	0.00	3	1.60	0	0.00
A→T	2	0.02	0	0.00	1	0.12	1	0.11	1	0.22	0	0.00	1	0.50	3	2.38	25	8.46	0	0.00	14	7.47	0	0.00
<i>A</i> → <i>T</i> at 686	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	1	0.50	1	0.79	6	2.03	0	0.00	8	4.27	0	0.00
$A \rightarrow T$ at OS	2	0.02	0	0.00	1	0.12	1	0.11	1	0.22	0	0.00	0	0.00	2	1.58	19	6.43	0	0.00	6	3.20	0	0.00
T→A	1	0.01	3	0.04	4	0.47	0	0.00	0	0.00	4	0.81	2	1.00	4	3.17	3	1.02	7	2.69	2	1.07	6	4.47
G→C	1	0.01	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	1	0.79	0	0.00	0	0.00	0	0.00	4	2.98
C→G	0	0.00	0	0.00	1	0.12	1	0.11	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
Indels	9	0.10	8	0.11	8	0.95	4	0.45	7	1.54	6	1.22	4	2.01	2	1.58	2	0.68	0	0.00	11	5.87	5	3.73
ΔΑ	3	0.03	2	0.03	3	0.36	1	0.11	1	0.22	2	0.41	0	0.00	2	1.58	1	0.34	0	0.00	1	0.53	0	0.00
∆A at 174-178	0	0.00	0	0.00	2	0.24	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
ΔA at OS	3	0.03	2	0.03	1	0.12	1	0.11	1	0.22	2	0.41	0	0.00	2	1.58	1	0.34	0	0.00	1	0.53	0	0.00
ΔΤ	1	0.01	2	0.03	2	0.24	0	0.00	5	1.10	1	0.20	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	1	0.75
ΔT at 201-205	0	0.00	0	0.00	1	0.12	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
ΔT at 255-260	0	0.00	1	0.01	1	0.12	0	0.00	2	0.44	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	1	0.75
∆T at OS	1	0.01	1	0.01	0	0.00	0	0.00	3	0.66	1	0.20	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
other single deletions	0	0.00	0	0.00	0	0.00	0	0.00	1	0.22	1	0.20	3	1.51	0	0.00	0	0.00	0	0.00	2	1.07	3	2.24
≥2 deletions	2	0.02	3	0.04	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
single insertions	3	0.03	1	0.01	3	0.36	3	0.34	0	0.00	2	0.41	1	0.50	0	0.00	1	0.34	0	0.00	8	4.27	1	0.75
≥2 insertions	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00	0	0.00
TOTAL	166	1.78	132	1.88	125	14.83	126	14.31	176	38.71	172	34.96	140	70.26	131	103.75	154	52.11	103	39.56	166	88.58	138	102.86
95% CI		1.10 2.29		1.32 2.87		12.95 18.73		10.72 21.90		30.22 48.90		26.92 52.80		55.28 168.82		90.97 158.86		37.51 69.88		15.67 61.67		13.70 257.65		62.25 173.87

<sup>a</sup> Number of events identified for given classes.

<sup>b</sup> Mutation rates [5-FOA<sup>R</sup>×10<sup>-6</sup>] for specific mutation types are shown in boldface. <sup>c</sup> Specific hotspot positions in the *URA3* coding sequence are indicated.

<sup>d</sup> OS – Other Sites.

**Table S2.** The analysis of statistical significance for the mutation spectra is presented in Fig. 3. *p* values were calculated using Fisher's exact test.

T→C vs A→G G→A vs C→T G→T vs C→A	DPB2 POL3 <b>OR2</b>	dpb2-100 POL3 OR1	DPB2 pol3-L612M OR1	dpb2-100 pol3-L612M OR2
	0.0007	0.3776	0.0005	NDª
DPB2 POL3 OR1	<0.0001	0.1733	<0.0001	ND
	0.0017	0.3702	0.0005	ND
	0.0012	0.5869	ND	ND
dpb2-100 POL3 OR2	0.2286	<0.0001	ND	ND
	<0.0001	<0.0001	ND	ND
	0.0008	ND	<0.0001	<0.0001
DPB2 pol3-L612M OR2	<0.0001	ND	<0.0001	<0.0001
	<0.0001	ND	<0.0001	<b>&lt;0.0001</b>
	ND	ND	>0.9999	<0.0001
dpb2-100 pol3-L612M OR1	ND	ND	<0.0001	<0.0001
	ND	ND	<0.0001	<0.0001

<sup>a</sup> ND – not determined

**Table S3.** The analysis of statistical significance for data showing the contribution of substitutions at specific hotspots to the total mutagenesis is presented in Fig. 4. *p* values were calculated using Fisher's exact test.

T→C at 97 <sup>a</sup> C→T at 310 G→A at 764 G→T at 679/706	DPB2 POL3 <b>OR2</b>	dpb2-100 POL3 OR1	DPB2 pol3-L612M OR1	dpb2-100 pol3-L612M OR2
	<b>0.0175</b> [1] <sup>b</sup>	0.0006 ↓ <sup>c</sup>	<b>0.0002</b> ↑	ND <sup>d</sup>
	<b>0.0169</b> [2]	0.3048	<b>0.0253</b> ↓	ND
DF BZ F OES OKT	<b>&lt;0.0001</b> [1]	<b>0.0407</b> ↓	<b>&lt;0.0001</b> ↑	ND
	<b>&lt;0.0001</b> [2]	0.8534	<0.0001 ↓	ND
	0.3489	0.0600	ND	ND
dab2 100 BOL 2 082	0.8899	<b>&lt;0.0001</b> [2]	ND	ND
apbz-100 POL3 OR2	0.7597	<b>&lt;0.0001</b> [1]	ND	ND
	<0.0001 ↓	<b>&lt;0.0001</b> [2]	ND	ND
	0.0508	ND	<b>&lt;0.0001</b> [1]	<b>&lt;0.0001</b> ↑
	<b>&lt;0.0001</b> ↑	ND	<b>&lt;0.0001</b> [2]	<b>&lt;0.0001</b> ↑
DPB2 pois-Lo 1210 OR2	>0.9999	ND	<b>&lt;0.0001</b> [1]	<b>&lt;0.0001</b> ↑
	<0.0001 ↓	ND	<b>&lt;0.0001</b> [2]	0.0004 👃
	ND	ND	<b>&lt;0.0001</b> ↓	<b>&lt;0.0001</b> [1]
dab2 100 ac12 / 612M OB1	ND	ND	<b>&lt;0.0001</b> ↑	<b>&lt;0.0001</b> [2]
upbz-100 p013-L012M OR I	ND	ND	<0.0001 ↓	<0.0001 [1]
	ND	ND	<b>&lt;0.0001</b> ↑	<b>&lt;0.0001</b> [2]

<sup>a</sup> Substitutions at specific hotspots are color-coded.

<sup>b</sup> The URA3 orientation with a higher contribution of substitutions at specific hotspots is shown in brackets: [1] – OR1. [2] – OR2.

<sup>c</sup> For *dpb2-100* and *pol3-L612M* mutants. an increase ↑ or decrease ↓ of substitutions at specific hotspots compared with the *DPB2 POL3* strain. is shown; for *dpb2-100 pol3-L612M* mutants. an increase ↑ or decrease ↓ of substitutions at specific hotspots compared with the *pol3-L612M* strain. is shown.

<sup>d</sup> ND – not determined

**Table S4.** The analysis of statistical significance for data showing the contribution of specific substitutions to the total mutagenesis is presented in Fig. 4. *p* values were calculated using Fisher's exact test.

T→Cª C→T G→A G→T	DPB2 POL3 <b>OR2</b>	dpb2-100 POL3 OR1	DPB2 pol3-L612M OR1	dpb2-100 pol3-L612M OR2
	<b>0.0006</b> [1] <sup>b</sup>	0.0294 ↓ <sup>c</sup>	<b>&lt;0.0001</b> ↑	ND <sup>d</sup>
	0.0928	0.0688	<b>&lt;0.0001</b> ↓	ND
DFB2 F0L3 OKT	<b>&lt;0.0001</b> [1]	0.7469	<b>&lt;0.0001</b> ↑	ND
	<b>&lt;0.0001</b> [2]	0.8290	<0.0001 ↓	ND
	<b>&lt;0.0001</b> ↑	<b>&lt;0.0001</b> [2]	ND	ND
dab2-100 BOL 2 082	<b>0.0160</b> ↑	<b>&lt;0.0001</b> [2]	ND	ND
<b>upb2-100</b> P 013 <b>UR2</b>	<b>0.0008</b> ↑	<b>&lt;0.0001</b> [1]	ND	ND
	<b>0.0007</b> ↓	<b>&lt;0.0001</b> [2]	ND	ND
	<b>0.0098</b> ↓	ND	<b>&lt;0.0001</b> [1]	<b>&lt;0.0001</b> ↑
DPP2 pol2 / 612M OP2	<b>&lt;0.0001</b> ↑	ND	<b>&lt;0.0001</b> [2]	<b>&lt;0.0001</b> ↓
	0.2544	ND	<b>&lt;0.0001</b> [1]	<b>&lt;0.0001</b> ↑
	<b>&lt;0.0001</b> ↓	ND	<b>&lt;0.0001</b> [2]	<b>&lt;0.0001</b> ↓
	ND	ND	<b>&lt;0.0001</b> ↓	<b>&lt;0.0001</b> [1]
dph2 100 pol2 / 612M OP1	ND	ND	<b>&lt;0.0001</b> ↑	<b>&lt;0.0001</b> [2]
ap52-100 p013-201210 OR1	ND	ND	<b>&lt;0.0001</b> ↓	<b>&lt;0.0001</b> [1]
	ND	ND	<b>&lt;0.0001</b> ↑	<b>&lt;0.0001</b> [2]

<sup>a</sup> Specific substitution types are color-coded.

<sup>b</sup> The URA3 orientation with a higher contribution of specific substitutions is shown in brackets: [1] – OR1. [2] – OR2.

<sup>c</sup> For *dpb2-100* and *pol3-L612M* mutants. an increase ↑ or decrease ↓ of specific substitutions compared with the *DPB2 POL3* strain. is shown; for *dpb2-100 pol3-L612M* mutants. an increase ↑ or decrease ↓ of specific substitutions compared with the *pol3-L612M* strain is shown.

<sup>d</sup> ND – not determined

Table S5. The analysis of statistical significance for the mutation spectra is presented in Fig. 6. p values						
were calculated using Fisher's e	exact test.					
	DPB2	dpb2-100	DPB2	dpb2-100		

T→A vs A→T	DPB2 POL2 <b>OR2</b>	dpb2-100 POL2 OR1	DPB2 pol2- M644G OR1	dpb2-100 pol2-M644G OR2
DPB2 POL2 OR1	0.1429	0.1251	0.2913	NDª
dpb2-100 POL2 OR2	0.0007	<0.0001	ND	ND
DPB2 pol2-M644G OR2	>0.9999	ND	<0.0001	>0.9999
dpb2-100 pol2-M644G OR1	ND	ND	0.2691	<0.0001
<sup>a</sup> ND – not determined				

Table S6. Yeast strains used in this work.

Strain	Relevant genotype	Source
YTAK001	agp1::URA3-OR1	[43]
Y485-3	agp1::URA3-OR1 rev3∆ DPB2 msh6∆	[44]
Y485-4	agp1::URA3-OR1 rev3∆ DPB2 msh6∆	[44]
Y773	agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y774	agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y771	agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y772	agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
YTAK002	agp1::URA3-OR2	[43]
Y486-2	agp1::URA3-OR2 rev3∆ DPB2 msh6∆	[44]
Y486-5	agp1::URA3-OR2 rev3∆ DPB2 msh6∆	[44]
Y776	agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y777	agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y778	agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y775	agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y779	agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
SNM12	pol3L612M agp1::URA3-OR1	[90]
Y491-3	pol3L612M agp1::URA3-OR1 rev3Δ DPB2 msh6Δ	[44]
Y492-1	pol3L612M agp1::URA3-OR1 rev3Δ DPB2 msh6Δ	[44]
Y783	pol3L612M agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y780	pol3L612M agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y781	pol3L612M agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
Y782	pol3L612M agp1::URA3-OR1 rev3∆ dpb2-100 msh6∆	This work
SNM24	pol3L612M agp1::URA3-OR2	[90]
Y493-1	pol3L612M agp1::URA3-OR2 rev3Δ DPB2 msh6Δ	[44]
Y494-1	pol3L612M agp1::URA3-OR2 rev3Δ DPB2 msh6Δ	[44]
Y786	pol3L612M agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y787	pol3L612M agp1::URA3-OR2 rev3∆ dpb2-100 msh6∆	This work
Y785	pol3L612M agp1::URA3-OR2 rev3Δ dpb2-100 msh6Δ	This work
Y784	pol3L612M agp1::URA3-OR2 rev3Δ dpb2-100 msh6Δ	This work
Y788	pol3L612M agp1::URA3-OR2 rev3Δ dpb2-100 msh6Δ	This work
SNM70	pol2M644G agp1::URA3-OR1	[59]
Y487-2	pol2M644G agp1::URA3-OR1 rev3Δ DPB2 msh6Δ	[50]
Y488-1	pol2M644G agp1::URA3-OR1 rev3A DPB2 msh6A	[50]
Y804	pol2M644G agp1::URA3-OR1 rev3Δ dpb2-100 msh6Δ	This work
Y805	pol2M644G agp1::URA3-OR1 rev3Δ dpb2-100 msn6Δ	
Y806	po/2M644G agp1::URA3-OR1 rev3Δ dpb2-100 msh6Δ	This work
1807	pol2M044G agp1::URA3-UR1 rev3Δ apb2-100 msrioΔ	
1000	poi2M644G agp1::URA3-URT rev3d apb2-100 msn6d	
1809	pol2M644G agp1::URA3-URT rev3d apb2-100 msn6d	This work
1010 SNM70	poi2M644G agp1URAS-ORT 16V5D 0pb2-100 115116D	
V180-2	p0/2/1/044G dyp1URA3-UR2 po/2/1/644G orp1://IPA2.OP2.rev24.DPR2.msh64	[59]
1409-2 V400-1	pol2M644G agp1URA3-OR2 16V3D DFD2 Institud	[50]
V701	pol2N644G agp 1 ORAS OR 2 revise DF D2 make A	
V702	pol2M644G agp1URAS-OR2 IEVSD upb2-100 INSN0D	This work
V703	pol2M644G agp1:.URA3-OR2 rev3A dpb2-100 msh6A	This work
Y794	poi2iniorred agp10145-012 rev3d apb2-100 msh6A	This work
Y795	poi2M644G agp1::01043 012 rovod upb2 rov manod	
SC228	MATa CAN1 his7-2 leu2-A"kanMX4 ura3-A tro1-289 ade2-1 lvs2-AGG2899-2900 DPR2	[25 39]
SC234	MATa CAN1 his7-2 leu2-Δ::kanMX4 ura3-Δ trp1-289 ade2-1 lys2-ΔGG2899-2900 dpb2-100	[25,39]

## Table S7. Primers used in this study.

Primer	Sequence 5'-3'
Rev3_UPTEF	CAATACAAAACTACAAGTTGTGGCGAAATAAAATGTTTGGAAATGAGATCTGTTTAGCTTGCC
Rev3_DNTEF	ATAACTACTCATCATTTTGCGAGACATATCTGTGTCTAGATTATTCGAGCTCGTTTTCGACAC
msh6UTEF	CAGATAAGATTTTTAATTGGAGCAACTAGTTAATTTTGACAAAGCCAATTTGAACTCCAAAAGATCTGTTTAGCTTGCC
msh6DTEF	CAACGACCAAAACTTTAAAAAAAAAAAAAAAAAAAAAATCTTACATACATCGTAAATGAAAAATATTCGAGCTCGTTTTCGACAC
Rev3-R4	TGACCACTCACATGGCGCTTTG
Rev3A	AATTCTGCCAATCTATTTGATCTTG
nat1UO	ACCGGTAAGCCGTGTCGTCAAG
Rev3-F4	AAAGGGCGAGCACAACTACTAC
Rev3D	CACCAGATAGAGTTTTGAACGAAAT
nat1DO	GCTTCGTGGTCGTCTCGTACTC
MSH6-UO	TAAAGTCGCTGGAGTAGG
msh6up2	GAATCCTTGGAGGAAGAC
HPH-UO	ACAGACGTCGCGGTGAGTTCAG
MSH6-DO	TCAAGCACCATCCTCAAG
msh6dw2	CCCATTCTTGCCCAAGATGC
HPH-DO	TCGCCGATAGTGGAAACCGACG
URA3F393	AACGAAGGAAGGAGCACAGAC
URA3R412	CCGAAATTCCTGGGTAATAAC
LEUBamF	AGTGGATCCACATACCTAATATTATTGCC
LEUMvaR	AAGGAGCATTCTGACAGAGTAAAATTCTTGAGGG

Impairment of the non-catalytic subunit Dpb2 of DNA Pol  $\epsilon$  results in increased involvement of Pol  $\delta$  on the leading strand Dmowski et al., https://doi.org/10.1016/j.dnarep.2023.103541 SUPPLEMENTARY MATERIAL Mva1269 Mva1269 LEU2 LEU2 Hind III. **BamHI** HindIII. ´BamHI ~BET2 -BET2 BspEl BspEl pLD2 pLD2-100 8810 bps 8810 bps DPB2 dpb2-100 ori pMB1 ori pMB1 ola YPR1740 PR1740 <mark>Pstl</mark> Xhol stl Xhol Scal Scal HindIII HindIII pAG29 pMJDPB2 pKF107 pJK1 PCR-amplified fragment

**Figure S1.** Maps of plasmid pLD2 and pLD2-100. For their construction. plasmids pAG29 [91]. pMJDPB2 [26]. pKF107 [25] and pJK1 [25] were used. The PCR fragment containing the *LEU2* gene was amplified using primers LEUBamF and LEUMvaR (Table 7). Restriction sites used to generate the gene cassette for yeast transformation are indicated by yellow backlight.

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