

Supplementary data

Supplementary data

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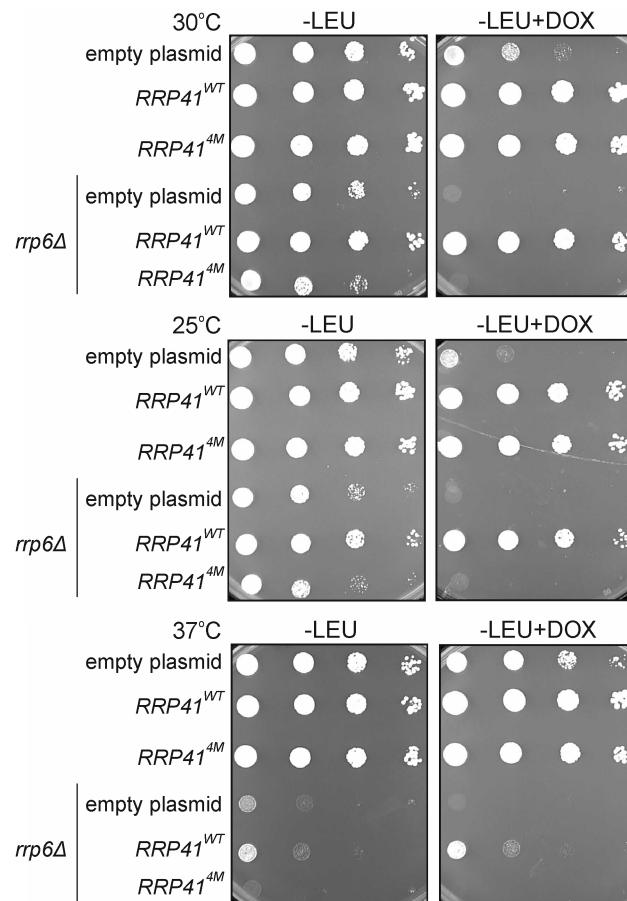
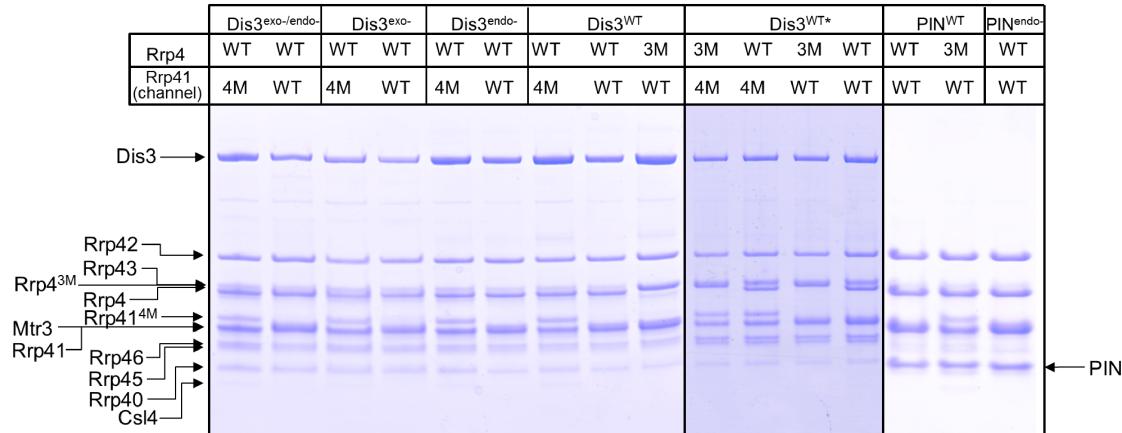


Figure S1. *Rrp41^{4M}* exosome central channel mutation is synthetically lethal with deletion of *RRP6*. LEU-marked plasmids containing *RRP41^{WT}*, *RRP41^{4M}* or no insert (empty plasmid control) were transformed into two strains harboring endogenous *RRP41* under control of a doxycycline-repressible promoter, the second was also an *rrp6* deletion strain. Growth phenotypes of the resulting strains were analyzed after 60 h at 25, 30 and 37°C in the absence (-LEU) or presence (-LEU+DOX) of doxycycline.

A



B

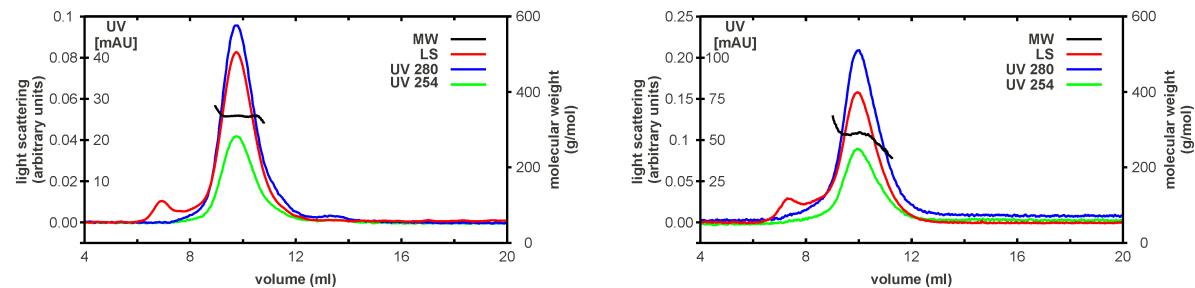


Figure S2. Reconstruction of WT and various mutant forms of the exosome complex from *C. thermophilum*. **(A)** SDS-PAGE analysis of reconstructed complexes. **(B)** Size exclusion chromatography combined with Multi Angle Light Scattering (SEC-MALS) analysis of reconstructed WT exosome complex. Black, red, blue and green lines represent calculated molecular weight, scattering signal, 280 nm and 254 nm absorbance respectively.

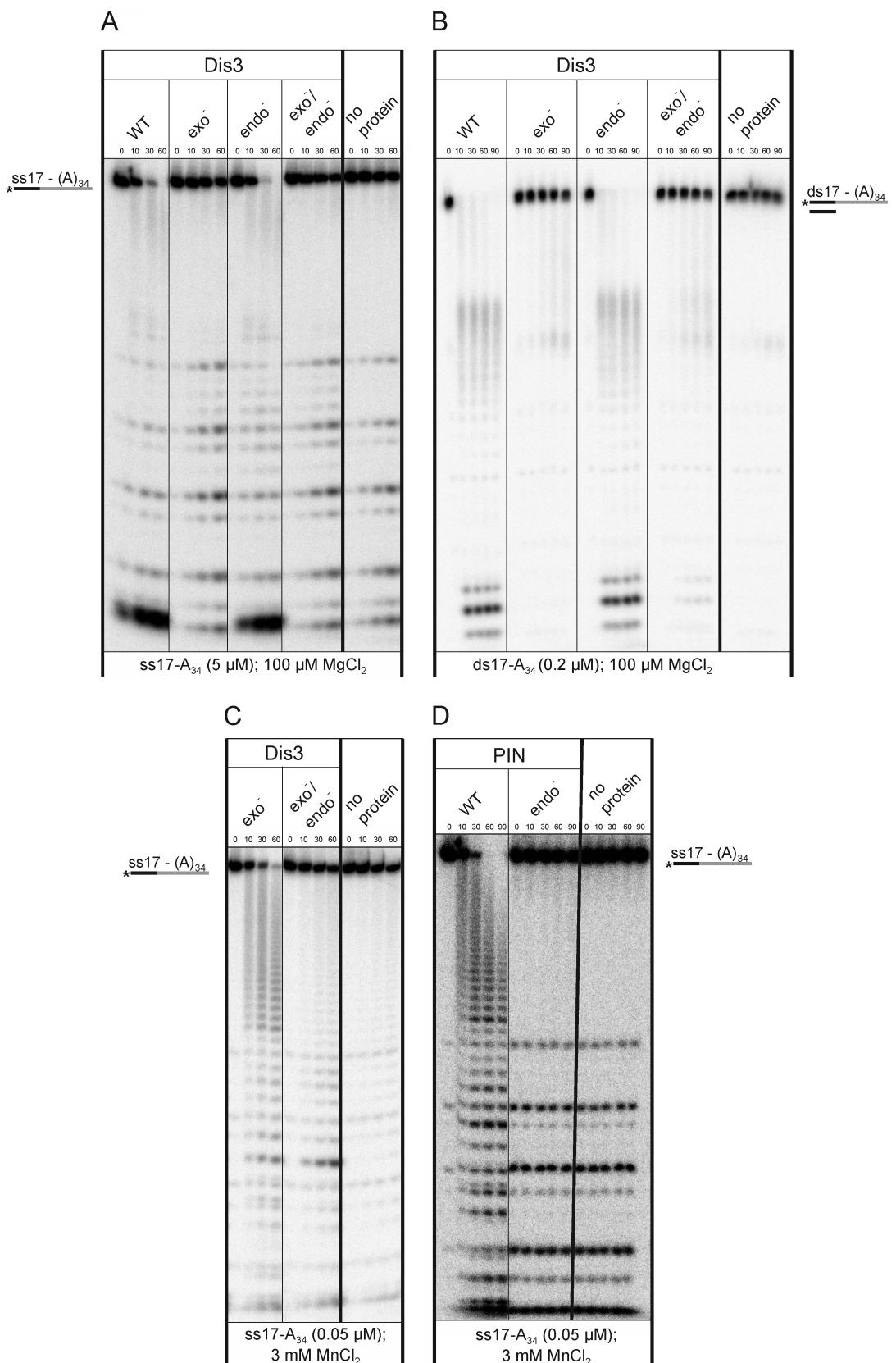


Figure S3. Biochemical characterization of *C. thermophilum* Dis3 ribonucleolytic activities. (A) and (B) Recombinant full-length *C. thermophilum* Dis3 protein displays exoribonucleolytic activity able to degrade both single- and double-stranded RNA substrates, which is strictly dependent on the intact aspartic acid residue in the active site of the RNB domain. 5'-labeled

ss17-(A)₃₄ (**A**) or ds17-(A)₃₄ (**B**) substrate was incubated in buffer containing 100 μM magnesium with different versions of Dis3 protein from *C. thermophilum* (Dis3^{WT}, Dis3^{exo-}, Dis3^{endo-}, Dis3^{exo-/endo-}) or in the absence of added protein. Samples were collected at the indicated time points (minutes) and analyzed by denaturing PAGE and phosphorimaging. (**C**) and (**D**) Recombinant full-length *C. thermophilum* Dis3 protein and its PIN domain alone both possess endoribonucleolytic activity, which is abolished by mutation of the conserved aspartate within the catalytic center of the PIN domain. 5'-labeled ss17-(A)₃₄ substrate was incubated in buffer containing 3 mM manganese with different versions of full-length *C. thermophilum* Dis3 (Dis3^{exo-}, Dis3^{exo-/endo-}) (**C**), isolated PIN domain (PIN^{WT}, PIN^{endo-}) (**D**) or in the absence of added protein. Samples were collected at the indicated time points (minutes) and analyzed by denaturing PAGE and phosphorimaging.

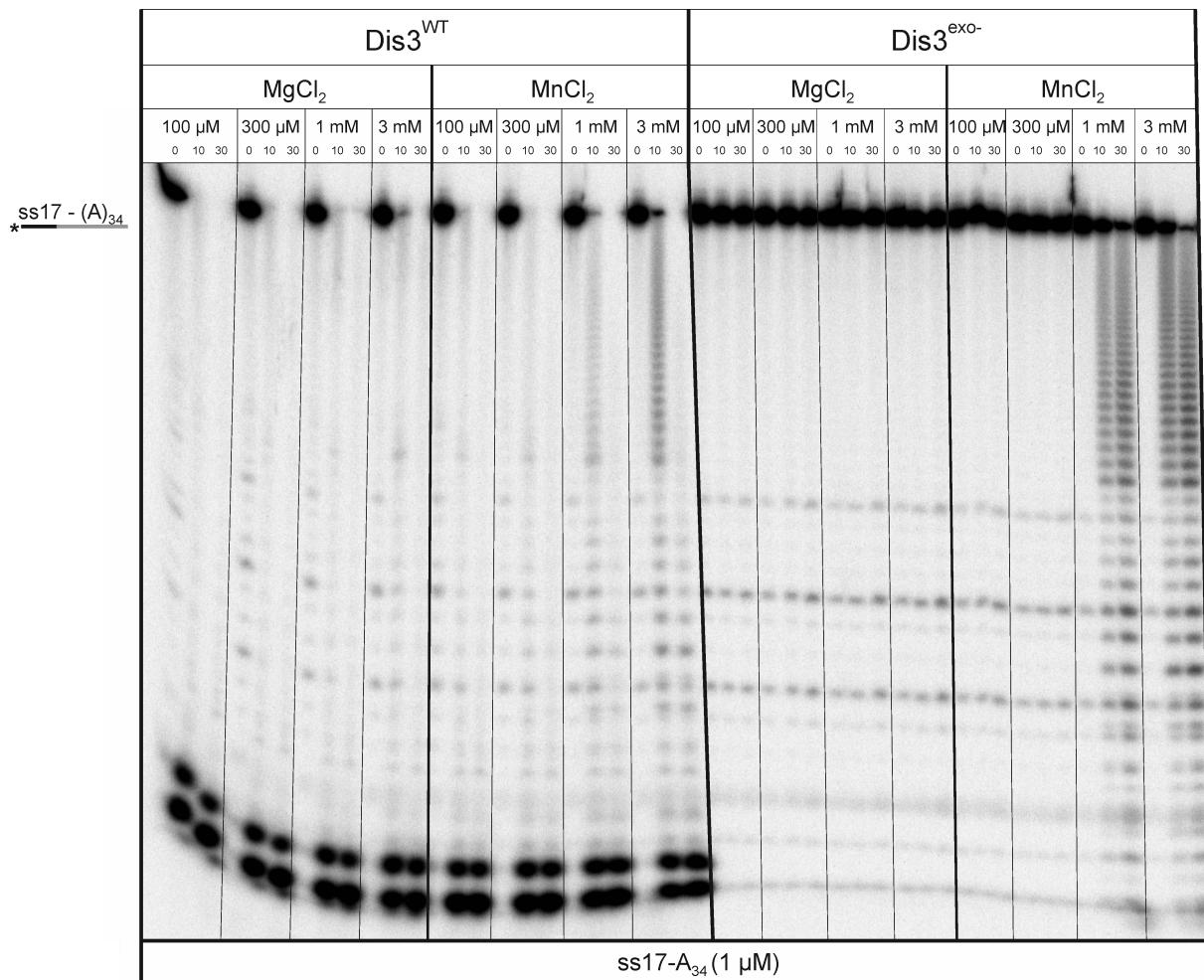


Figure S4. *C. thermophilum* Dis3 exoribonucleolytic activity can be observed within a broad range of both magnesium and manganese concentrations while PIN domain-dependent endoribonuclease activity is detectable exclusively in buffers containing manganese at a high concentration. 5'-labeled ss17-(A)₃₄ RNA substrate was incubated with *C. thermophilum* Dis3^{WT} or Dis3^{exo-} in buffers containing magnesium or manganese cations at different concentrations as indicated. Reactions were terminated at the indicated time points (minutes) followed by denaturing PAGE and phosphorimaging. Dis3^{WT} displays mainly exoribonuclease activity, which is readily visible in all cases while Dis3^{exo-} endoribonuclease activity is strictly dependent on the presence of manganese as a cofactor and can be revealed only when the concentration of this cation is at least 1 mM.

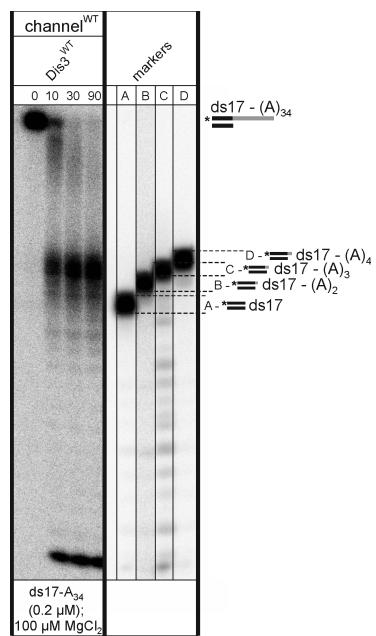


Figure S5. Degradation of double-stranded ds17-(A)₃₄ RNA by *in vitro* reconstructed *C. thermophilum* exosome results in accumulation of decay intermediates possessing 2-3 nt-long single-stranded extensions. ds17-(A)₃₄ substrate was incubated in buffer containing 100 μ M magnesium with reconstituted exosome containing non-mutated channel and Dis3^{WT}. Samples were collected at the indicated time points (minutes) and analyzed by denaturing PAGE and phosphorimaging. Blunt-ended dsRNA (ds17) or its counterparts with short single-stranded extensions (ds17-(A)₂, ds17-(A)₃ and ds17-(A)₄) were run in parallel, as indicated in the right, to enable estimation of the size of decay intermediates.

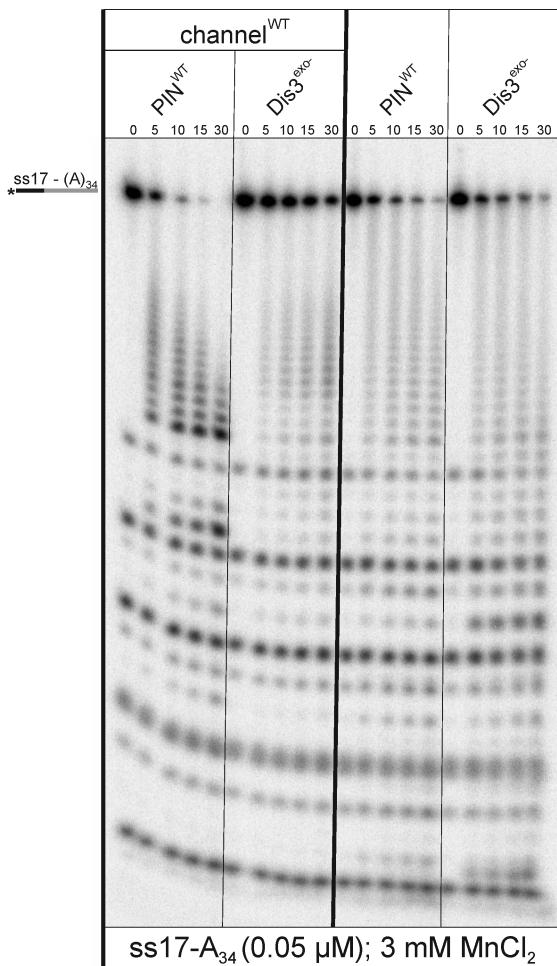


Figure S6. Direct comparison of degradation rates and patterns performed for reconstituted exosomes and individual proteins that display endoribonucleolytic activity. 5'-labeled ss17-(A)₃₄ substrate was incubated in a buffer containing 3 mM manganese with *C. thermophilum* exosome complexes with PIN domain (PIN^{WT}) or full-length Dis3 lacking exonuclease activity (Dis3^{exo}) as well as with PIN^{WT} and Dis3^{exo} proteins alone. Samples were collected at the indicated time points (minutes) and analyzed by denaturing PAGE and phosphorimaging.

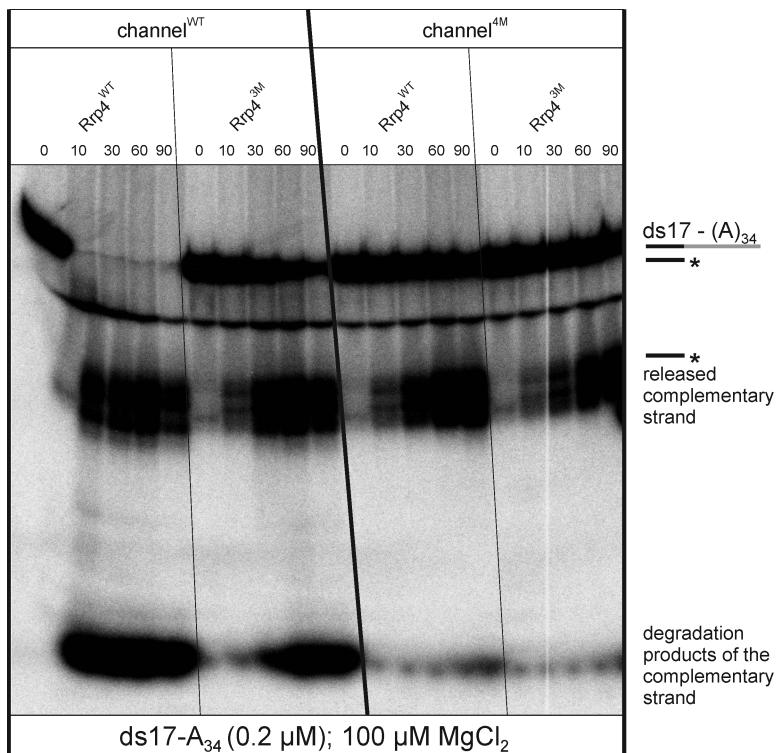


Figure S7. Unwinding of dsRNA substrate by reconstructed *C. thermophilum* exosome complexes. Reactions were carried out using ds17-(A)₃₄ duplex RNA substrate, which had been prepared in the presence of 5'-labeled complementary ss17 oligonucleotide and incubated with *in vitro* reconstituted *C. thermophilum* exosomes containing different versions of the channel (either with Rrp41^{WT} or Rrp41^{4M}) and Rrp4 cap subunit (Rrp4^{WT} or Rrp4^{3M}), as indicated. Samples were collected at the indicated time points (minutes) and analyzed by electrophoresis in 15% native polyacrylamide gel, followed by phosphorimaging. Positions of the initial duplex RNA substrate, the radioactive complementary strand released by unwinding of dsRNA and the final degradation products of the complementary strand are shown in the right.

Table S1. Oligonucleotides used for strain construction, northern blot analysis and biochemical assays. “r” before sequence in brackets indicates that the oligo is composed of ribonucleotides.

Oligonucleotide	Sequence
ADZKD100	TTAATAGTAGTCTATGCTGG
ADZKD103	CTGTTTTATGTACTTTATATAAACAGTGGCAATTAAATGGCG TTTTTATTttaaaactggatggcgccgt
ADZKD106	TCGGTCATATGAGAGTGTGTTGCG
ADZKD107	AGTGGTTAGTGGTAAAATCCAACGTTGCCATCGTTGGGCC CCGGTCG
ADZKD113	TAGACGAAATAGGAACAAACAAACAGCTTATAAGCACCCAATAA GTGCGTTtagatctgttagcttgcc
ADZKD114	ATGAAAATTACCATATTTATAAAATAAAAAAAATACGCTTGTTT ACATAAtcgatgaattcgagctg
ADZKD117	TTTGATCCCGTAGAACACG
ADZKD118	TGGCTTGGTGGTGTGTTGC
5S rRNA probe	CTACTCGGTCAAGGCTC
7S rRNA probe	GGCCAGCAATTCAAGTTA
5.8S rRNA probe	GCGTTGTTCATCGATGC
SCR1 probe	ATCCCCGGCCGCCTCCATCAC
ss17	r(CCCCACCACCAUCACUU)
ss17-(A) ₂	r(CCCCACCACCAUCACUUAA)
ss17-(A) ₃	r(CCCCACCACCAUCACUUAAA)
ss17-(A) ₄	r(CCCCACCACCAUCACUUAAAA)
ss17-(A) ₁₄	r(CCCCACCACCAUCACUUAAAAAAAAAAAAAA)
ss17-(A) ₃₄	r(CCCCACCACCAUCACUUAAAAAAAAAAAAAA AAAAAA)
compl	r(AAGUGAUGGUGGUGGGG)

Table S2. Yeast strains employed in this study.

Strain	Mutations	References
ADZY522	<i>rrp41</i> ^{4M} (K62E S63D R95E R96E)	This study
ADZY531	<i>dis3</i> ^{endo-} (D171N)	This study
ADZY537	<i>rrp41</i> ^{4M} (K62E S63D R95E R96E) <i>dis3</i> ^{endo-} (D171N)	This study
BSY1735	<i>dis3</i> ^{exo-} (D551N)	(1)
ADZY539	<i>rrp41</i> ^{4M} (K62E S63D R95E R96E) <i>dis3</i> ^{exo-} (D551N)	This study
BSY1665	<i>dcp</i> ¹⁻²	(1)
ADZY649	<i>dcp</i> ¹⁻² <i>rrp41</i> ^{4M} (K62E S63D R95E R96E)	This study
ADZY712	<i>RRP4 tet-off mtr4gfp</i>	This study
TH_6151	<i>RRP4 tet-off</i>	(2)
TH_3687	<i>RRP41 tet-off</i>	(2)
ADZY524	<i>RRP41 tet-off rrp6Δ</i>	This study

Table S3. Constructs used for *C. thermophilum* exosome reconstruction.

Construct	Plasmid backbone
HisTAG-SUMO-CtRrp46 (WT) + CtRrp43 (WT)	pET28
HisTAG-SUMO-CtRrp41 (WT) + CtRrp45 (WT)	pET28
HisTAG-SUMO-CtRrp41 (R62E R63D K114E R115E) + CtRrp45 (WT)	pET28
HisTAG-SUMO-CtMtr3 (WT)	pET28
HisTAG-SUMO-CtRrp42 (WT)	pET28
HisTAG-SUMO-CtRrp4 (WT)	pET28
HisTAG-SUMO-CtRrp4 (R158E K159D R160D)	pET28
HisTAG-SUMO-CtCsl4 (WT)	pET28
HisTAG-SUMO-CtRrp40 (WT)	pET28
HisTAG-SUMO-CtDis3 (WT)	pET28
HisTAG-SUMO-CtDis3 (D168N)	pET28
HisTAG-SUMO-CtDis3 (D536N)	pET28
HisTAG-SUMO-CtDis3 (D168N D536N)	pET28
HisTAG-SUMO-CtPIN (WT)	pET28
HisTAG-SUMO-CtPIN (D168N)	pET28

Rrp42

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Ch._thermophilum
Consensus_ss:

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Ch._thermophilum
Consensus_ss:

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Ch._thermophilum
Consensus ss:

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Ch._thermophilum
Consensus ss:

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Ch._thermophilum
Consensus ss:

Conservation:
RRP42_Homo_sapiens
Rrp42p_S._cerevisiae
Rrp42p_Glycine_max

Mtr3

MTR3
Conservation:
MTR3_Homo_sapiens
Mtr3p_Ch._thermophilum
Mtr3p_S._cerevisiae
Consensus ss:

Conservation:
MTR3_Homo_sapiens
Mtr3p_Ch._thermophilum
Mtr3p_S._cerevisiae
Consensus ss:

Conservation:
MTR3_Homo_sapiens
Mtr3p_Ch._thermophilum
Mtr3p_S._cerevisiae
Consensus sequence:

Conservation:
MTR3_Homo_sapiens
Mtr3p_Ch._thermophilum
Mtr3p_S._cerevisiae
Conservation score:

Conservation:
MTR3_Homo_sapiens
Mtr3p_Ch._thermophilum
Mtr3p_S._cerevisiae
Conservation score:

Rrp43

Rrp43
Conservation:
RRP43_Homo_sapiens
Rrp43p_Ch._thermophilum
Rrp43p_S._cerevisiae
Consensus ss:

Conservation:
RRP43_Homo_sapiens
Rrp43p_Ch._thermophilum
Rrp43p_S._cerevisiae
Consensus ss:

Conservation:
RRP43_Homo_sapiens
Rrp43p_Ch._thermophilum
Rrp43p_S._cerevisiae
Consensus sequence:

Conservation:
RRP43_Homo_sapiens
Rrp43p_Ch._thermophilum
Rrp43p_S._cerevisiae

Conservation:		9	
RRP43_Homo_sapiens	174	TALAEVN-----LKKKSYLNIRTHPVATSFAVFD-----	202
Rrp43p_Ch._thermophilum	248	REMVCS-----KTETMKLTIKGLPIACSAAVFLEKE-----HGEVA	284
Rrp43p_S._cerevisiae	252	ASDLRMTI RTRGRSATIRETYEIIICDQTKSPLMINAKNIAFSASN GIVEVLDPECQLQNSDNSEEVEVDI	321
Consensus_ss:	eeeeee	eeeeeeeeeee	

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Conservation:
RRP43_Homo_sapiens          263 KKLMLDEVIKSMKPK  276
Rrp43p_Ch_.thermophilum     353 KEVMK-----  357
Rrp43p_S_.cerevisiae       389 STRFNI-----  394
Consensus ss:                hhhh

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Rrp46

Conservation:
RRP46_Homo_sapiens -----
Rrp46p_S._cerevisiae -----
Rrp46p_Ch._thermophilum 254 LHWKS 258
S. pombe

Rrp40

RRP40		99	999
Conservation:			
RRP40_Homo_sapiens	1 MAEPASVAAESLAGSRARA ARTV LQGVVLPGEELLPEQEDEAEGPGGAVER PLS LNARACSRVR VVC GPG		
Rrp40p_Ch._thermophilum	1 MSTT----- TRP FVLPG E TIDPSLV-----		
Rrp40p_S._cerevisiae	1 ----- MSTF1 FPGDSFVFDP-----		
			70
			33
			24

Conservation:		9 9	9 9 9	9 9 9	9	
Rrp40_Homo_sapiens	71	LRRCG--DRLLVTKCGRRLRHKEPGSNSGGGVWVDSQQKRRYVPVKGD HIVIGIVTAKSGDIFKVDVGGS --				137
Rrp40p_Ch._thermophilum	34	LRHVPP-SDI IPTVAGQLITNLN----KNSM WVEYN SQRVPTQND LVLAQVLRSTQDSYLCLITPHTP				97
Rrp40p_S._cerevisiae	25	IYCDPNTQEIRPVNTGVHLVS AKGK-SGVQTAYID YSSKR YIPSVD FVIGVIIGTFSDSYKVSLQN FSS				93

Conservation:		9 99		9 9 99 99	
RRP40_Homo_sapiens	207	IRKLLAPD-----CEIIQEVGKLYPLEIVFGMNGRIVWAKTIQQTLLILANILEACEH-			259
RRP40p_Ch._thermophilum	168	ARRLLMAKSREEGKVGVLEMLAGEDPSIGEAGLAGFETAVGRNGRVWVGSEDVKTVIIVGRALQETDRG			237
RRP40p_S._cerevisiae	163	ARQLLFNND-----FPLLKVLAAHKTFEVAIGLNGKIWVKCEE	LSNTLACYRTIMECCQK		217

Conservation:		9	
RRP40_Homo_sapiens	260	-MTSDQRKQIFSRLAES-----	275
Rrp40p_Ch._thermophilum	238	NLTIEGQRKLVRLRLEMR-----	256
Rrp40p_S._cerevisiae	218	-NDTAAFKDIAKRFKEILTVKEE	240

Supplementary materials and methods

Strain construction

rrp41^{4M} mutant

Mutations K62E S63D R95E R96E were introduced into the *RRP41* gene by *in vivo* recombination into a BMA64 wild type diploid strain. A DNA fragment containing the *RRP41* gene with K62E S63D R95E R96E mutations and Sp.*HIS5* selection marker was amplified using ADZKD100 and ADZKD101 primers in overlap PCR with two templates: first – *RRP41* gene with K62E S63D R95E R96E mutations amplified on the pRS415 plasmid with the cloned *RRP41* gene containing K62E S63D R95E R96E mutations (pADZ433), second – Sp.*HIS5* gene amplified on pFA6A-link-yEGFP-Sp.*HIS5* vector. Spores were dissected and those harboring *RRP41* mutations yielded strain ADZY522.

dis3^{endo-} mutant

The D171N mutation was introduced into the *DIS3* gene in the BMA64 wild type haploid strain by *in vivo* recombination. The *DIS3* gene with the D171N mutation, protein A sequence and KITRP1 selection marker was amplified using ADZKD102 and ADZKD103 primers and pBS3278 vector as a template. This gave strain ADZY531.

rrp41^{4M} dis3^{endo-} mutant

The ADZY531 strain was modified by introducing *RRP41* K62E S63D R95E R96E mutations and Sp.*HIS5* selection marker by *in vivo* recombination using a DNA fragment amplified from ADZY522 total DNA as a template using ADZKD100 and ADZKD101 primers. This gave the ADZY537 strain.

rrp41^{4M} dis3^{exo-} mutant

A DNA fragment containing the *RRP41* gene with K62E S63D R95E R96E mutations and Sp.*HIS5* selection marker was amplified using primers ADZKD100 and ADZKD101 with ADZY522 total DNA as a template and introduced into the BSY1735 (1) strain by *in vivo* recombination. This gave strain ADZY539.

dcp¹⁻² rrp41^{4M} mutant

A DNA fragment containing the *RRP41* gene with K62E S63D R95E R96E mutations and Sp.*HIS5* selection marker was amplified using primers ADZKD100 and ADZKD101 with ADZY522 total DNA as a template and introduced into the BSY1665 (1) strain by *in vivo* recombination. This gave strain ADZY649.

The strain was transformed with pRP485 (containing *MFA2pG*), pRP603 (containing wild type *PGK1pG*), pRP611 (containing *PGK1pG* with PTC), pRP1079 (containing nonstop *PGK1pG*) and pRP1252 (containing *PGK1pG* with stem loops).

RRP41 tet-off *rrp6Δ*

A DNA fragment containing Sp.*HIS5* selection marker and sequences flanking the *RRP6* gene was amplified using primers ADZKD113 and ADZKD114 with pFA6A-link-yEGFP-Sp.*HIS5* vector as a template and introduced into the endogenous *RRP6* locus in the TH_3687 strain (2). This gave strain ADZY524.

RRP4 tet-off Mtr4gfp

A DNA fragment containing the *MTR4* gene with gfp sequence and Sp*HIS3MX6* selection marker was amplified using ADZKD117 and ADZKD118 primers with total DNA from a yeast strain with Mtr4 gfp-tagged (3) as a template and introduced into the TH_6151 strain (2) with the *RRP4* gene under the control of the tet-off promoter by *in vivo* recombination. This gave strain ADZY712.

Supplementary references

1. Dziembowski,A., Lorentzen,E., Conti,E. and Seraphin,B. (2007) A single subunit, Dis3, is essentially responsible for yeast exosome core activity. *Nat. Struct. Mol. Biol.*, **14**, 15-22.
2. Mnaimneh,S., Davierwala,A.P., Haynes,J., Moffat,J., Peng,W.T., Zhang,W., Yang, X., Pootoolal,J., Chua,G., Lopez,A. et al. (2004) Exploration of essential gene functions via titratable promoter alleles. *Cell*, **118**, 31-44.
3. Huh,W.K., Falvo,J.V., Gerke,L.C., Carroll,A.S., Howson,R.W., Weissman,J.S. and O'Shea,E.K. (2003) Global analysis of protein localization in budding yeast. *Nature*, **425**, 686-691.